



Published in final edited form as:

Perception. 2008 ; 37(10): 1529–1559.

Visual Prosthesis

Peter H. Schiller and Edward J. Tehovnik

Abstract

There are more than 40 million blind individuals in the world whose plight would be greatly ameliorated by creating a visual prosthetic. We begin by outlining the basic operational characteristics of the visual system as this knowledge is essential for producing a prosthetic device based on electrical stimulation through arrays of implanted electrodes. We then list a series of tenets that we believe need to be followed in this effort. Central among these is our belief that the initial research in this area, which is in its infancy, should first be carried out in animals. We suggest that implantation of area V1 holds high promise as the area is of a large volume and can therefore accommodate extensive electrode arrays. We then proceed to consider coding operations that can effectively convert visual images viewed by a camera to stimulate electrode arrays to yield visual impressions that can provide shape, motion and depth information. We advocate experimental work that mimics electrical stimulation effects non-invasively in sighted human subjects using a camera from which visual images are converted into displays on a monitor akin to those created by electrical stimulation.

1. Introduction

When Pope Benedict XII set out to have the walls of the great cathedral of St. Peter redecorated in the 12th century, he sent messengers all over Italy to obtain samples from the best artists. When a messenger came to Ambrogio Bondone Giotto (1267–1337), who had not heard of the contest, he did not have a sample painting prepared. So he took a sheet of paper and a pencil dipped into red ink, and drew a perfect circle. “Here is your drawing,” he said. The Pope, upon examining all the productions submitted, some of which were quite fancy and elaborate, chose Giotto. To this day in Tuscany there is the saying “The round O of Giotto” (Broos, 1971).

Turning this story around, we can pose the question central in considering prosthetic devices for the blind: how is it possible for us to identify a perfect circle? Answers to this question, which relate to how the brain processes shape information, are central to the goal of creating a functional prosthetic device for the blind.

In the United State there are over one million blind individuals (Leonard, 2002). In the world the number exceeds 40 million. A variety of strategies have emerged to aid the blind, which involve both invasive and noninvasive approaches. Creating devices that can convey visual impressions through non-visual modalities is one of the more successful strategies, one outcome of which is the ability of blind individuals to read using the somatosensory system by palpating raised dots that form shapes of letters as in the case of Braille as devised by Louis Braille in 1881 (Mellor, 2006). In this paper we shall consider some effective methods that would recreate visual images by virtue electrical stimulation of the visual system. In doing so three central questions need to be posed:

- A. What aspects of vision need to be reproduced by a prosthetic device?
- B. Which animal species are the most suitable for testing?
- C. Which visual areas are best for such implantation?

The first step in developing a prosthesis based on electrical stimulation is to have a thorough understanding of the workings of the visual system. A prosthetic device has to operate in harmony with the workings of this system. The next section provides a brief discussion of the visual system.

2. The basics of the visual system

a. Retina

Five general classes of neurons have been identified in the retina. The first set consists of the photoreceptors. Using the Golgi method it was discovered that they form two major subclasses in the primate retina, the rods and the cones (Schultze, 1873). Rods, which are absent in the fovea, are for night vision and cones for day vision. The diameter of cones in the fovea in monkeys and humans are about 2.4 μm but become progressively larger and with more space between them to accommodate the rods as depicted in Figure 1A. In the foveal area there are approximately 200,000 cones per square millimeter which drops ten fold at an eccentricity of 5 degrees from the fovea (Barlow, 1981; Rodieck, 1973).

The second set of retinal cells are the horizontal cells, the third the bipolar cells, the fourth the amacrine cells and the fifth the ganglion cells. The ganglion cells send their axons to the central nervous system. Their axons course along the inner surface of the retina as can be seen in Figure 1C and exit from the eye at the optic disk that appears in Figure 1B and C. The information from the rods and cones, where they coexist on the retina, converges on the ganglion cells through different retinal pathways.

The photoreceptors in the retina are placed closest to the pigment epithelium that is right against the inner wall of the sclera. The other retinal elements are positioned more centrally in the eye. Consequently, the photons entering the eye have to travel through the cells and fibers to activate the photoreceptor molecules. To minimize interference with the incoming photons, these cells and axons are transparent. To further improve visual acuity in the fovea the cells and their processes are placed radially sideways, thereby making for clear access of light to the foveal photoreceptors. This arrangement is shown in Figure 1D. Because of this and because of the high density of the photoreceptors, the retinal ganglion cells that receive inputs from the foveal receptors are stacked on top of one another in regions beyond the foveal area thereby forming several sub-layers as shown in Figure 1D; with increasing eccentricities and the concomitant decrease in the density of the photoreceptors, eventually the ganglion cells are reduced to a single sublayer.

In characterizing the structure and function of the neurons in the retina, methods were developed to record intracellularly from retinal cells and, once functionally characterized, labeled by injecting dyes into the cells (Werblin and Dowling, 1969). This work established the following: (1) all photoreceptors hyperpolarize to light and that they produce only graded potentials; the neurotransmitter in the receptors is glutamate (Bloomfield and Dowling, 1987). (2) Horizontal cells also all hyperpolarize and have only graded potentials. (3) Some bipolar cells hyperpolarize and some depolarize; they all produce only graded potentials. (4) Some of the amacrine cells produce only graded potentials, but some also generate action potentials. (5) Retinal ganglion cells, whose axons exit from the eye to central brain structures, yield action potentials.

It has been shown that there are several different classes of retinal ganglion cells that perform different analyses of the visual scene. Notable among these are the ON and OFF retinal ganglion cells discovered by Keffer Hartline in the 1930s. He showed that some cells discharged when light was shone into their receptive fields and some discharged when the light was terminated. He therefore called them the ON and OFF retinal ganglion cells. The ON and OFF systems

arise at the level of the cone bipolar cells by virtue of bipolar cells that have either sign-conserving or sign-inverting neurotransmitter receptor sites. The OFF bipolar cells have non-NMDA receptor sites and hyperpolarize to light just as do the photoreceptors. The ON bipolars, on the other hand, have a different neurotransmitter receptor called mGluR6 that inverts the signal; when the photoreceptors hyperpolarize, the ON bipolar cells depolarize (Rodieck, 1973; Schiller, 1996). These cells therefore depolarize when the photoreceptors hyperpolarize. The ON and OFF bipolar cells connect to the ON and OFF ganglion cells. As a consequence of this arrangement, an excitatory signal in the form of action potentials can be generated for light decrement in OFF ganglion cells and for light increment in ON ganglion cells.

In many primates the rod photoreceptors connect with a separate class of bipolar cells, the rod bipolars, which all have neurotransmitter receptors that are sign conserving. In the rod system these bipolar cells connect to the same ganglion cells as do the cones. They do this through a series of steps that involves amacrine cells and yields sign conserving and sign inverting signals for the ganglion cells. Thus under daylight conditions the ganglion cells are driven predominantly by the cones whereas at night they are driven by the rods in those portions of the retina where the rods and cones coexist. Thus it appears that there are no separate pathways to the brain for rods and cones.

A small population of retinal ganglion cells receive convergent input from the ON and OFF bipolar cells (Hartline, 1938; Schiller, 1996; Wässle and Boycott, 1991). These cells make extensive projections to the superior colliculus (Schiller and Malpeli, 1977)

As we move our eyes about in exploring the visual scene, with each shift in gaze there is either an increment or decrement in illumination at any single point on the retinal surface. The ON cells discharge when there is an increment in illumination and the OFF cells discharge when there is a decrement. Light increment and decrement are mutually exclusive; as we move our eyes about exploring a pattern-rich visual scene, changes in illumination at any point on the retinal surface will be either incremental or decremental. It has been discovered that the ON and OFF systems in the retina actually provide independent coverage of the visual field. Accordingly, in the inner retina the dendritic processes of the ON and OFF cells form separate sub layers (Wässle and Boycott, 1991).

The ability to process chrominance cues in humans and many primates is accomplished by having three kinds of cones, those that are most sensitive to short wavelengths (blue), medium wavelengths (green) and long wavelengths (red).

The receptive fields of retinal ganglion cells have circular center/surround organization (Kuffler, 1953). The surround is inhibitory. Consequently a small spot of light (or darkness) placed into the center of the receptive field of ganglion cells produces a vigorous response, whereas a large spot produces greatly attenuated activity. The manner in which ON and OFF retinal ganglion cells discharge to light incremental and light decremental spots is shown schematically in Figure 2A. In the upper portion of the figure small spots confined to the excitatory center of the receptive fields are used. The ON cell shown discharges vigorously to the bright spot, whereas the OFF cell discharges to the dark spot. When large spots, which act on both the center and the antagonistic surround of the receptive fields, are used instead, the response is much attenuated.

One reason why retinal ganglion cells have developed this antagonistic center/surround organization is that this arrangement optimizes sensitivity; the cells discharge to *changes* in contrast, not to absolute levels of illumination (Rodieck, 1973; Schiller, 1996). Retinal ganglion cells have a relatively limited range of discharge rates from 0 to about 400Hz. But illumination levels vary over almost 10+ log units and the pupil diameter can change only over roughly a 16-fold range in humans and most primates. Hence adaptation, much of which takes place in

the photoreceptors themselves, is of central importance. Most retinal ganglion cells cease to respond at any level of maintained light. Thus it appears that these types of retinal ganglion cells can best be described as *local illumination change detectors*.

This view is supported by experiments in which images were stabilized on the retina (Pritchard, 1961). One approach was to attach a tiny projector to a contact lens which subjects were asked to wear while lying down. It was found that images disappeared in a matter of just a few seconds when they were “fixed” on the retina. This can actually be demonstrated in a simpler way using gaussian stimuli that lack sharp edges, so the normal slight tremor of the eye, which keeps images alive on the retina, is not a factor. An example of this is shown in Figure 2B. If one fixates on the upper central square spot for about 10–30 seconds, both gaussian smudges disappear because the retinal ganglion cells adapt and become silent. If one now shifts the center of gaze to the lower fixation spot, complementary afterimages appear; the area of the retina that had been exposed to the dark gaussian on the right now yields the afterimage of a light spot by virtue of the discharge of ON ganglion cells. The reason this happens is that, due to adaptation during the first fixation, the region exposed to the dark gaussian becomes more sensitive so that after shifting the gaze the photons reflected from the gray background impinge on this more sensitive region, activating ON-center ganglion cells and thereby creating the complementary afterimage. The opposite situation arises for the area exposed to the bright gaussian during initial fixation; the portion of the retina exposed to this bright spot becomes less sensitive. Consequently, after the gaze shift the photons from the gray background activate a less sensitive region of the retina, which in turn activates the OFF cells yielding the percept of a (non-existing) dark spot.

The apparent size of afterimages changes as a function of viewing distance. To experience this, fixate on the little white center square within the black disk in Figure 2C with the figure at a comfortable viewing distance. Maintain rigorous fixation for about 30 seconds and then shift your gaze to the cross in the right of Figure 2C. As soon as the negative afterimage appears, move your head toward or away from the page. The size of the afterimage will change as a function of viewing distance; the closer your head to the page, the smaller the afterimage becomes. A more dramatic way of perceiving this size change is to reestablish the afterimage and then look at a clear region on the wall in the room. The image will appear to be quite large. The scaling of the afterimage is fully lawful.

These observations raise the point that under normal viewing conditions we have a very strong sense of size constancy. The perceived size of a quarter, for example, remains the same irrespective of the distance at which it is viewed, although the size of its image on the retina changes lawfully with viewing distance. Information available in the brain regarding the accommodative state of the lens, the degree of vergence of the two eyes and object familiarity are important here.

Another important factor of note, however obvious, is that while normal objects in the visual scene appear stationary when the eye moves about, afterimages move with the eyes. Thus the information available in the brain regarding the position of the eye in orbit, the state of vergence of the eyes, and the accommodative state of the lens play a central role in enabling us to determine the location of objects in three-dimensional space relative to our bodies and to perceive a stable world.

Further work examining the retina has established that there are several additional classes of ganglion cells that deal with different aspects of visual analysis (Schiller & Logothetis, 1990; Schiller et al., 1990a,b; Wässle and Boycott, 1991). Most studied among these in the primate are the so called midget and parasol retinal ganglion cells. The midget cells are small and have medium conduction velocities to the central nervous system. The parasol cells, which are less

numerous, have dendritic arbors that look like umbrellas. The midget system can process fine detail as well as color. Cells of the parasol system have much larger receptive fields, are very sensitive to changes in luminance contrast, and play an important role in motion processing. They respond transiently to illumination changes, producing brief bursts of action potentials. Midget cells produce more sustained activity, but following each illumination change their activity returns to baseline in a few seconds. Overall, the receptive fields of ganglion cells increase in size with increasing eccentricity from the fovea, with the parasol cells having receptive sizes three times the diameter of midget cells (Rodieck, 1973). The midget and parasol cells both come in ON and OFF sub-varieties. The midget cells also process color in separate subclasses; the receptive fields of these cells are tiny, with the center in most composed of but a single cone. By contrast, the parasol cells receive a mixed input from the three cone types both in the center and surround regions of their receptive fields; as a result, this highly sensitive system can respond to color exchanges but cannot specify what the colors are.

Several other classes of retinal ganglion cells have been identified. These include the cells of Dogiel, which are directionally selective. They form part of the accessory optic system (AOS) which ties in with the vestibular and oculomotor systems and plays a central role in stabilizing the eye with respect to the world (Karten et al., 1977; Simpson et al., 1979) and cells that provide information about overall levels of illumination (Barlow and Levick, 1969).

Given these various classes of retinal ganglion cells that perform largely different jobs in analyzing the visual scene, one can appreciate the fact that creating a prosthetic device that can activate them selectively is a daunting task. If a prosthetic device were to be based on electrical stimulation of the retina, most of these different classes of cells likely would be activated indiscriminately with largely unknown sensory and motor consequences.

Several other problems must be overcome for a retinal prosthetic device to become workable. A central one of these is the fact that there are a million fibers coursing to the optic disk where the retinal ganglion cells exit the eye. As a result, there is a tightly packed plexus of fibers on the inner surface of the retina as shown in Figure 1. Were these fibers activated by electrical stimulation through a metal microelectrode with its tip on the inner retinal surface, the receptive fields of the activated ganglion cells of origin would form a banana-like swath (Schiller and Malpeli, 1977). This fact is best realized by the finding that microstimulation of the retina produces phosphenes whose characteristics could not be predicted by simply noting the site of stimulation within the retina (Rizzo et al., 2003a,b; but see Humayun et al., 1999). This results when the stimulation activates fibers-of-passage and when the position of the electrode within the substrate is not stable. Even if these problems were to be solved (Humayun et al., 2003), several other concerns would need to be addressed. Given that there are several distinct classes of retinal ganglion cells that perform different analyses on the visual scene, their indiscriminate activation would likely yield confusing precepts and undesirable motor acts such as eye movements generated by the cells of Dogiel and pupillary changes (Simpson et al., 1979).

The density of the receptors and ganglion cells changes dramatically with eccentricity (Rodieck, 1973). In central retina, ganglion cells can be stacked on top of each other radially five deep, as already noted, whereas in the periphery they form just a single layer. Selective activation of ganglion cells in the foveal representation is therefore a problem. Also, selective activation of ganglion cells would require extremely tightly spaced stimulating electrodes with the capacity of delivering different current levels for different locations. Also problematic is the fact that the movement of our eyes is taken into account by the accessory optic system as well as the vestibular system; the information so provided keeps track of the position of the eye in orbit that thereby enables us to localize objects in space relative to our bodies. It is doubtful that electrical stimulation of the retina could provide this information. This problem

is also present when the lateral geniculate nucleus (LGN) and area V1 are electrically stimulated; we shall discuss this in more detail below.

Yet another problem with using a retinal prosthetic is that, in many cases, the retinae of blind individuals undergo dramatic changes over time. The retina can degenerate to a level that forecloses the effective stimulation of neurons (Bartlett et al., 2005; Santos et al., 1997; Stone et al., 1992).

Degeneration is also of concern in the LGN. Several studies have established that in this structure significant changes occur after deprivation and enucleation (Hendry, 1991; Matthews et al., 1960; Mize et al., 1992; Sloper et al., 1987a, b; Tigges and Tigges, 1991). By contrast, studies have established that area V1 remains an effective site for eliciting phosphenes by electrical stimulation in individuals who have been blind for many years (e.g. Dobelle et al., 1976; Schmidt et al., 1996).

b. The central projections from the retina

With the eyes moving to the front of the head in many species, including primates, the connections from the retina changed dramatically as depicted in Figure 3. With the eyes focusing on a central spot, the visual field can be divided into two halves. The right visual field projects to the left hemisphere and the left visual field to the right. This is accomplished by having the temporal hemiretinae project ipsilaterally and the nasal hemiretinae project contralaterally. This arrangement allows local visual activation from punctate locations in space in each eye to end up at corresponding locations in the brain thereby preserving the spatial integrity of the system.

The projections from the various cell types in the retina have considerable order. The cells of Dogiel project to the so-called terminal nuclei, and yet another class of cells, often referred to as w-cells, project extensively to the superior colliculus (Rodieck, 1973; Schiller and Malpeli, 1977; Simpson et al., 1979). The most robust projection in the primate, however, is to the lateral geniculate nucleus of the thalamus (LGN), a cross section of which appears in Figure 4A. The anterior portion of the LGN has six layers as shown in this figure. Beyond approximately 17 degrees of eccentricity the LGN is reduced to four layers (Malpeli and Baker, 1975). Overall the LGN is a rather small structure measuring about 6 millimeters antero-posteriorly as well as dorsoventrally and 5 millimeters laterally in the monkey.

The LGN is a laminated structure in which there is an orderly topography. The upper, parvocellular layers, receive input from the midget cells whereas the lower, magnocellular layers receive input from the parasol cells. In central retina and in the central representation of the visual field in the LGN, the midget cells outnumber the parasol cells approximately 8 to 1; in peripheral representation their respective numerosity becomes nearly equal (Malpeli and Baker, 1975). Yet another group of cells, the koniocellular cells, project to the intralaminar layers, some of which are believed to play a role in processing information from the blue cones (Chatterjee and Callaway, 2003; Martin et al., 1997). The input from the left and right eyes is layer specific. The arrangement of the connections is such that the left LGN process information from the right half of the visual field with the obverse being the case for the right LGN.

The receptive field properties of LGN cells in the monkey are quite similar to those found in the retina (Schiller and Malpeli, 1977; Wiesel and Hubel, 1966). Due to the much higher packing density of retinal ganglion cells in central vision, a higher volume of tissue in the LGN is devoted to central vision.

At this stage we have only limited knowledge of what visual percepts are created by electrical stimulation of the LGN as few studies have been conducted on humans (Marg and Dierssen,

1965), but some recent work is ongoing in animals (Pezaris and Reid, 2007). Marg and Dierssen (1965) found that stimulation of the LGN evoked spots and disks of light exhibiting particular colors of blue, green, yellow, or red. Lined visual streaks were also elicited. This might be related to the fact that fibers of passage between and LGN and cortex were activated and that the eyes of the subjects were not controlled. Much like the retina, the receptive fields of the cells in the LGN are anchored to the eyes. Marg and Dierssen (1965) found that stimulation of the LGN produced a strong desire to move the eyes in the direction of the evoked percept.

Animal studies have shown that electrical microstimulation of the LGN for the most part produces a punctate percept (Pezaris and Reid 2006, 2007). The topographic arrangement of the LGN makes it feasible but difficult to place arrays of electrodes in an orderly fashion into this structure. Although the volume of tissue in the LGN is significantly greater than in the retina, the relatively small overall size of this structure, its location deep in the brain, and the laminar separation of cell types raises concerns about the feasibility of proper electrode array placement (Cohen, 2007). Selective activation of different classes of cells is quite difficult as is the orderly placement of electrodes that can preserve the spatial integrity of the system. Worrisome also is the stability that can be achieved when such long electrodes are used and the possible damage they might cause in the overlying tissue.

c. The striate cortex (V1)

In primates the information provided by the retinal ganglion cells and the cells of the LGN is transformed in several notable ways in area V1 as first established by Hubel and Wiesel (1977). The majority of cells in V1 are orientation and direction specific. Many receive convergent input from the midget and parasol ON and OFF cells (Schiller et al., 1976a,b). Cells also receive an orderly, convergent input from the two eyes that gives rise to stereoscopic depth perception (Barlow et al., 1967; Freeman and Ohzawa, 1990; Poggio and Fischer, 1977).

In their early work, Hubel and Wiesel identified two major classes of cells in area V1 which they called simple and complex (2005). Complex cells receive convergent input from ON and OFF cells that originate in the retina. Because of this convergence, these cells respond to both light increment and decrement and hence seem unable to specify sign of contrast. The majority of simple cells also receive convergent input from the ON and OFF systems, but a subgroup of them does receive selective input from either the ON or the OFF cells that project from the LGN to V1. Yet another group of cells in V1 responds in a selective and tonic fashion to either light increment or light decrement, variously called T-cells or luxotonic cells (Bartlett and Doty, 1974; Kayama et al., 1979; Schiller et al., 1976 a, b). Thus the subgroup of simple cells and the T-cells have the attributes necessary to specify the sign of contrast of objects that appear in their receptive fields. Since these various groups of cells are largely intermingled in V1, they cannot be selectively stimulated using methods we presently have at our disposal.

The gray matter in the striate cortex is relatively uniform, measuring about 2 millimeters in thickness as shown in Figure 4B. The packing density of cells is high and is also quite uniform throughout area V1 (O’Kusky and Colonnier, 1982; Peters, 1987, 1994; Rockel et al., 1980). This is not true for the retina and LGN. For the retina in particular, there is a higher density of cells for central vision as compared to peripheral vision and the overall density of cells is less when compared to area V1 (Barlow, 1981; Perry and Cowey, 1985; Schein, 1988). This difference has serious implications for the use of electrical stimulation: a non-uniform density means that different current levels would be needed to be used to activate a fixed number of cells (Tehovnik, 1996). This is not a major problem for area V1 since a fixed current will activate a fixed number of cells throughout; the delivery of 2.0 μA of current is estimated to have a spread of 50 μm in V1 (Tehovnik and Slocum, 2007a).

In the rhesus monkey the central 6–8 degrees of the visual field is laid out on the surface of the occipital lobe. This region for the most part is lissencephalic meaning that it is quite smooth with the exception of the shallow *external calcarine fissure* as shown in Figure 4D. Also shown is the topographic layout of this region with much more area allocated to central than peripheral representation of the visual field. The projection is such that the images on the cortical surface of the monkey are actually upside down, with the lower visual field represented in the upper portion of area V1 and the upper visual field represented in the lower portion. As already noted, the left visual field projects to the right hemisphere and the right visual field to the left. The sizes of the receptive fields in the fovea are very small (< 0.5 degrees) and increase with increasing eccentricity (Dow et al., 1981).

The gray matter of V1 has both a laminar and columnar organization. Inputs from the LGN terminate most extensively in layer 4C, with layer 4C β receiving input mostly from the magnocellular layers whose input from the retina is from the parasol cells, whereas the input to layer 4C β is from the parvocellular layers of the LGN to which the midget cells project from the retina (Fitzpatrick et al., 1985; Hubel and Wiesel, 1972; LeVay et al., 1975). The koniocellular cells, which from the retina project to the intralaminar layers of the LGN, terminate in the uppermost layers of V1 (Hendry and Yoshioka, 1994).

In area V1 the two major kinds of columns distinguished are orientation and the alternating left and right ocular inputs, which are well defined in layer 4 (Hubel and Wiesel, 1977). The columns representing the left and right eyes alternate so that when this input is labeled from one of the eyes, which yields dark stripes, a top view of the cortex gives the appearance of zebra stripes. This orderly separation of left and right eye inputs is most evident in layer 4. In the upper and lower layers the majority of cells receive a convergent input from the two eyes thereby integrating their input. One aspect of this integration involves the processing of stereoscopic depth information. The columnar organization as seen in layer 4 looks much like a thumb imprint. Interestingly, the width of the columns is constant, about 500 μm , in spite of the fact that progressively less volume of tissue is allocated to increasing eccentricities of visual field representation.

The organization of area V1, its sizable volume (e.g. about 2000 mm^3 per hemisphere—Felleman and van Essen, 1991) with over 100 times as many neurons as the LGN (Barlow, 1981), and the effects of local electrical stimulation, which creates a small image (Brindley and Lewin, 1968; Dobbelle and Mladejovsky, 1974; Schmidt, 1996; Tehovnik and Slocum, 2007a), suggest that this area is well suited for studying the feasibility of a prosthetic device. V1 in monkeys is immediately below the skull. Therefore, electrodes can be placed accurately and with relative ease (e.g. Bradley et al., 2005; Troyk et al., 2003). The large volume of tissue representing the visual field in area V1 is an important consideration. It allows for selective activation of tissue representing areas just a fraction of a degree apart, thereby allowing reasonable re-creation of shapes. The volume of gray matter allocated for spatial representation in V1 is over 30 times greater than in the LGN and over 100 times greater than in the retina (Felleman and van Essen, 1991; Perry and Cowey, 1985; Schein, 1988; Winter et al., 1969).

Most of the neurons of V1 respond relatively transiently to static changes in illumination as do the retinal ganglion cells (Hubel and Wiesel, 1977). The V1 cells can be thought of as local illumination difference detectors but with the additional restrictions pertaining to the orientation of edges, direction of motion and spatial frequency of objects in the visual scene.

Extensive work has been carried out in humans and to a lesser extent in monkeys showing that electrical stimulation of area V1 creates a punctate image much like a star (Brindley and Lewin, 1968; Dobbelle and Mladejovsky, 1974; Schmidt, 1996; Tehovnik and Slocum, 2007a). The size of the image varies with eccentricity, being very small in the fovea and growing in size

with increasing distance from central vision in accordance with magnification factor (Brindley and Lewin, 1968; Tehovnik and Slocum, 2007a). When a region in V1 coding for a visual-field eccentricity of 3 degrees was stimulated in a monkey, the contrast of the image created with 20–120 μA was 6 to 12% with size of 15 to 20 minutes of visual angle (Schiller et al., 2006).

In discussing the retina, we had noted that one of the major problems in this area is that the tissue degenerates over time (Matthews et al., 1960; Sloper et al., 1987a,b). In advocating area V1 as the target of a visual prosthesis, one needs to address to what extent this area undergoes changes after retinal damage. One set of studies examining this issue showed some significant changes in area V1 following damage to the retina (Chino et al., 1992; Darian-Smith and Gilbert, 1995; Gilbert et al., 1990; Gilbert and Wiesel, 1992; Heinen and Skavenski, 1991; Kaas et al., 1990). Another set of studies, however, showed little change in the organization of V1; particularly notable among these studies is the finding that humans blind for a decade or more experience phosphenes when their area V1 is electrically stimulated. (Dobelle et al., 1976; Schmidt et al., 1996; Smirnakis et al., 2005). These latter studies suggest that the stability of area V1 over time is significantly greater than what has been found for the retina and the LGN.

3. Processing of the visual scene in three dimensions

The visual scene projected into the eye forms a two dimensional image on the retina. One of the prime tasks of the visual system is to reconstruct the third dimension from this image. The mechanisms for depth processing include stereopsis, motion parallax, perspective and shading (Casagrande and Boyd, 1996; Howard, 2002; Roe et al., 2007). Stereopsis is achieved by having neurons in the cortex receive convergent input from the two eyes that enables the system to convert disparity cues into depth information. At this stage it is unlikely that a prosthetic device based on electrical stimulation can be devised that can selectively stimulate neurons in the visual system that specify different depths based on disparity. The representation of different depths does not appear to be arranged in columns or layers that could be selectively activated by electrical stimulation. While stereopsis is indeed a major mechanism for depth perception, it should be noted that 5 to 10% of the population in the USA is stereo blind. Such individuals rely on other depth cues the brain can process. Among these, motion parallax is perhaps the most prominent mechanism. This is a very robust system as deficits in this capacity are extremely rare.

Motion parallax is a monocular cue and takes advantage of the fact that when there is movement, objects at different distances from the observer move at different rates across the retinal surface. This differential rate of motion is processed by the brain to provide information about the relative location of objects in depth. A prosthetic device that can accurately activate successive elements in an array based on the visual input from a camera should be able to provide this information reasonably well. Other available depth cues, as we had noted in section 2a, include shading, perspective, vergence and accommodation.

4. Eye movements and spatial localization

The visual system as we had described so far, is organized in retinocentric coordinates. Such organization is well suited to represent what is often called allocentric space -- how objects in three-dimensional space are arranged relative to one other (Klatzky, 1998). An additional task for the brain is to compute where objects are located in space *relative* to the body, often referred to as egocentric space. This is an essential requirement, without which organisms that locomote in space would not be able to do so with any degree of accuracy. To properly orient in space, information about the position of the limbs and head relative to the body and the position of the eye in orbit need to be accurately computed and integrated.

The movement of the eyes and the signals needed to keep track of them, involves several rather elaborate processes in the brain. Since one of the prime tasks of eye movements is to be able to accurately direct the fovea to desired target locations so that targets can be analyzed in fine detail, the saccadic system, which accomplishes this for the most part, is coded in retinocentric coordinates. This is the case in several brain structures including V1, the superior colliculus and the frontal eye fields. Electrical stimulation at any given site in these areas elicits a saccadic eye movement that has a constant vector; the same saccadic vector is generated no matter where the eyes are in orbit (Schiller and Tehovnik, 2001). In V1 and the superior colliculus the vector depends on where within the structure one stimulates. If one first plots out the receptive field location of the neurons relative to the foveal area, their activation results in a saccadic eye movement that shifts the center of gaze into the receptive field. Further proof that these areas code constant vectors comes from the observation that prolonged electrical stimulation produces a staircase of saccades with similar successive vectors (Robinson, 1972; Schiller and Stryker, 1972).

To keep the eye stabilized with respect to the world we have the vestibulo-ocular reflex. If the head is rotated, the semicircular canals send a signal to the oculomotor complex that with great rapidity rotates the eye in the opposite direction to keep it stable with respect to the visual scene. Doing so is essential for being able to process visual information when we are in motion. Many of us have seen movies made with cameras attached to the head of skiers. The hills and trees vibrate wildly on the trip downhill and we are impressed by the incredible skiing abilities of these skiers. The fact is, however, that what the skiers see is nothing like what the camera sees. The skiers have a wonderful vestibular system that stabilizes the eye with respect to the ski slopes, whereas a regular camera does not have such a system. Indeed, people whose vestibular system is compromised have major difficulties in processing the visual scene when they are in motion (Leigh and Zee, 1980; Sherman and Keller, 1986).

The vestibulo-ocular reflex is readily modifiable, due apparently in part to the visual inputs from the accessory optic system we had already discussed (Simpson et al., 1979). When the body and head are rotated, the eye counter rotates. When this is done in most subjects the magnitude of the counter rotation of the eyes equals the body rotation. When this is measured quantitatively and performance is optimal, it can be said that the gain of the oculomotor reflex equals 1 (Jones, 1977; Jones et al., 1984). If one now proceeds to have a subject wear magnifying or minifying lenses for a few days, the gain of the reflex changes to accommodate to the change. If the gain is halved by the lens system, the vestibulo-ocular reflex changes to 0.5; if it is doubled, it changes to 2. It is believed that the visual input from the accessory optic system has the power to modify the gain, with the cerebellum playing a significant role in this process (Demer et al., 1985). The vestibulo-ocular reflex and its adaptability need to be integrated into the creation of a prosthetic device.

5. Phosphenes created by electrical stimulation and their relationship to eye movements

As we had already noted, electrical microstimulation of area V1 creates a small, star-like image. The eye, the LGN, and V1 utilize a retinocentric code as delineated in Figures 4, 5 and 6. The computation of the spatial location of visual images relative to the body, as described above, is accomplished by utilizing signals that define the location of the eyes in orbit, their vergence and the accommodation of the lens. The image created by electrical stimulation is not privy to this information. The phosphene moves with the eyes and changes in size as a function of vergence and accommodation (Brindley and Lewin, 1968; Cowey and Walsh, 2000; Dobbelle and Mladejovsky, 1974; Grèusser, 1991; Richards, 1971; Rushton and Brindley, 1977; Schmidt et al., 1996). The rules that govern the spatial location and size of the phosphene follow the same rules as do afterimages (Grèusser, 1991). For example: in sighted individuals, electrical

stimulation of area V1 coding for a region a few degrees from the fovea produces a star-like image with a diameter of approximately 0.2 degrees of visual angle. This means that when a subject views a sheet of paper at a distance of 57.3 centimeters from the eye, the star-like image has a diameter of 2 mm. This is equivalent to the lower case letter “a” using a size 12 Ariel font. As in the case of afterimages, doubling the distance of the sheet of paper from the eyes doubles the apparent size of the phosphene to 4 mm and halving the distance reduces its apparent size to 1 millimeter. Such size changes apply throughout, so that when an array of electrodes is activated, as described in Figures 7–10, the pattern produced remains the same but its overall apparent size and its location in space change as a function of eye movements, vergence and accommodation; the position of the phosphene relative to the fovea is constant and hence moves about when the eye moves just as do afterimages.

One major problem in creating an effective prosthetic device based on electrical microstimulation is that eye movements are compromised in most blind individuals. Eye movements are often uncoordinated, multidirectional, disjunctive, and the gain of the vestibulo-ocular reflex is less than 1. Some blind individuals have a persistent nystagmus. These conditions are more pronounced in the congenitally blind than in individuals who become blind later in life (Bartels, 1928; Hall and Ciuffreda, 2002; Kömpf and Piper, 1987; Leigh and Zee, 1980; Sherman and Keller, 1984). There are several possible solutions to this problem, depending to the nature of the eye-movement control problem in each individual, all of which necessitate bringing eye-movements under control. Some possible solutions will be discussed below.

6. The basic tenets for producing a visual prosthetic for the blind

In this section we list essential basic tenets that must be taken into account for the creation of an effective, long-lasting prosthetic device.

a. Must use an appropriate animal model

In setting up strategies for the research to examine this question, it is wise to take a close look at the development of the cochlear implant, which has been a tremendous success. Tens of thousands deaf people have cochlear implants that provide auditory information to the auditory centers of the brain (Clark, 2006). The multichannel auditory prosthetic device pioneered by Graeme Clark was developed over a period of many years starting in the late 60s. An effective device that was ready for implantation in human subjects occurred only after Clark and colleagues had performed ten years of animal experimentation perfecting their device by conducting electrophysiological and psychophysical experiments on behaving as well as anesthetized cats (Clark, 2003).

Following the strategy of the development of the cochlear prosthetic, it is our belief that the first step in developing a visual prosthetic for blind humans is to develop an animal model. This belief is not shared by all investigators; Weiland and Humayun (2003) have suggested that major device verification of effectiveness should be done on humans only. We believe that adopting this approach in the absence of animal experimentation will, at best, significantly delay progress; more likely, this approach will end in failure. So far, the field of visual prosthetics has opted for an approach that is antithetical to the one adopted by Graeme Clark. A recent example of this is the work of Ed Schmidt and colleagues who in 1996 tested the effects of stimulating through an array of electrodes implanted in V1 of a blind patient (Schmidt et al., 1996). This work ended abruptly due to complications (Wagenaar, 2004, pg. 4). The work was followed nine years later by the implantation in a monkey of a similar device that had previously been implanted in the blind patient (Bradley et al., 2005). Some five months after the electrodes had been implanted in the monkey the animal became lethargic due to fluid buildup around the electrodes. Following recovery, the animal was left with a persistent upward

nystagmus and visual field defects. Subjecting blind humans to an implant that has not been perfected in animals should be deemed unethical at this time. Nevertheless, humans should be used in entirely noninvasive procedures to test some of the basic assumptions and limitations of possible prosthetic devices before a visual prosthetic device has been perfected in animals.

b. Stimulation at each brain site must provide a punctate percept

As noted above, there is quite a bit of evidence, both in humans and monkeys, that electrical stimulation of area V1 produces a punctate, star-like image. The size of this image increases with increasing eccentricity of visual field representation in accordance with magnification factor (Tehovnik and Slocum, 2007a,b). This suggests that area V1 is a promising candidate for exploring the feasibility of a prosthetic device based on electrical stimulation. Extrastriate cortical areas appear to be less well suited for implantation of a prosthetic device for several reasons: (1) The receptive fields of single cells become progressively larger in the extrastriate areas (Felleman and Van Essen 1991); in V3 they are three times larger on the average; in V4 and MT they are much larger than that. (2) The topography in extrastriate areas is significantly less well defined than in V1. (3) The overall volume of tissue in most higher visual areas is less than in V1.

c. Must select a brain region in which a large area is devoted to visual processing

Area V1, as already noted, is quite uniform in thickness and cell density. The visual field is laid out in a neat topographic order and, in monkeys, the region representing the central 6–7 degrees of the visual field is on the lissencephalic cortical surface; this fact makes for relatively easy placement of electrode arrays. The study of other areas, notably the retina, the LGN, and higher cortical areas should continue to be explored, however.

d. Must preserve the spatial integrity of the system

To enable images to be identified, we believe that a central effort must be made to preserve the spatial integrity of the system. This concern brings us back to the Giotto story: How can we create a prosthetic device that, using appropriate electrical stimulation, can create images that correspond to real objects in the visual scene such as a circle? To deal with this question we need to take a closer look at the layout and topography of area V1 where the problem of preserving the spatial integrity of the system is more manageable than in the retina and the LGN.

To better understand how images are represented in area V1, a set of simple dotted images is shown in the upper portion of Figure 5 using the scheme of Schwartz (1994) who established that over short distances the cortical map is conformal within each hemisphere. Dots are used because, as already noted, electrical stimulation at each site in V1 produces a star-like image. The areas activated in area V1 of the monkey are depicted below in the figure. The images in the left visual field project to the right hemisphere and those in the right visual field to the left. The images are reversed and upside down on the retinal surface and on the surface of V1. Figure 5A shows how a set of dots forming an arrow placed in the center of the visual field is laid out in the cortex of the monkey. In Figure 5B a circular array of dots is presented in the right visual field. The areas activated in V1 form a roughly circular array with the area activated by each dot varying notably in size due to the magnification factor. Figure 5C shows what the activation is like when the identical circular array of dots is centered along the vertical meridian. The activation in the cortex now becomes bilateral. Being equidistant from the center point, which impinges on the center of the foveal representation, the size of each region activated in V1 is therefore the same. However, the activated region bears no resemblance to a circle; two curved lines are created. The equivalent percept, however, is a perfect circle. We believe that a prosthetic device must factor in this organizational principle if it is going to be effective for creating veridical visual images.

We propose that a proportional array, which creates a visual image of a rectangular group of dots when all elements are activated, is an advantageous arrangement. We shall present several lines of evidence to support this point. In Figure 6A a square array of 256 equally-spaced dots is shown, with 128 in the left and 128 in the right visual hemifield. The area of the visual field occupies 4 degrees of visual angle. The corresponding regions activated in area V1 of the monkey are shown schematically below. Were one to place an electrode at each of these locations in V1, as shown in the bottom of Figure 6B, the images created upon brief electrical stimulation of all 256 electrodes would presumably look like those shown on the top of Figure 6B. The overall layout of the square is similar to the image shown on the top of Figure 6A. However, due to the fact that the receptive-field size of the neurons increases with increasing eccentricity, as does the magnification factor, the dots created by the electrical stimulation would be smaller near the center than in the periphery. We shall call this arrangement the “proportional” array.

How effective such a device might be for pattern perception under static conditions is approximated in Figure 7. In Figure 7B a rear view of the monkey brain is shown with an array 128 electrodes placed into each hemisphere as had been described in Figure 6. Activating all these electrodes creates the image depicted in 7A. Figure 7C shows the visual field upon which the words FIAT LUX are flashed on briefly. The image in the camera is broken up into 256 sections with the aid of a computer, each of which is connected to a corresponding electrode. The connections are inverted and the right half of the visual field is connected with the electrode array on the left and the left visual field to the right, thereby preserving the integrity of the system. The arrows provide the layout of the connections. The electrodes activated by FIAT LUX are shown in red in 7E. The visual image assumed to be created appears in 7D. The words are reasonably readable.

Next, let us examine two other possible bilateral electrode arrays. Figure 8 shows a circular array, which is arranged according to the magnification factor in V1. Figure 8B shows the layout of the electrodes which when activated, creates the image shown in Figure 8A. The electrodes activated by the word FIAT LUX appear in red in 8C; the resultant image created is shown in 8D. The quality of this stationary image is quite good.

Let us now compare what kinds of images are created by electrical stimulation of an array of electrodes that are equally spaced as shown in Figure 9B. In this case each array consists of 256 elements, twice as many as those used in Figures 7 and 8. When all electrodes are activated a butterfly image is created as shown in 9A. Hence we shall refer to this arrangement as the “butterfly array.” The electrodes activated by FIAT LUX and the image created appears in 9E and 9D. The words are not nearly as readable as with the proportional arrays.

e. Must take into account the operation of the ON and OFF systems

The existence of the ON and OFF systems and their convergence onto single cells in area V1 raises some intriguing questions as to how to create a prosthetic device for the blind that can effectively process both light incremental and light decremental information.

As we had noted earlier, the ON and OFF systems have emerged over the course of evolution to enable living organisms to process both light incremental and light decremental information rapidly and efficiently. From the single-ended system of the photoreceptors, which all hyperpolarize to light, a double ended system has been created which yields largely separate excitatory signals for light increment (the ON system) and for light decrement (the OFF system). Notably, the majority of neurons in area V1 receive a convergent input from these two systems. Therefore, these cells do not seem to be able to provide viable information about the sign of contrast of objects in the visual scene.

We make about three saccades per second as a result of which images at specific retinal locations are present only for about a third of a second. Due to the relatively transient nature of the neural responses and the movement of the eyes, it may be said that with each shift in gaze the slate is wiped clean, ready to accept and analyze the image that falls on the retina during the subsequent fixation. Because of this state of affairs, signals from a putative prosthetic device using a camera, from which signals pass through a computer, need to be transient, thereby essentially mimicking the manner in which neurons are normally activated.

It is a well known fact that in blind subjects who lack a functional retina, electrical stimulation of individual sites in the LGN and V1 creates a star-like image on a black background. In such subjects stimulation does not appear to produce dark spots. It may be said then that electrical stimulation in the blind is single ended. The stimulation of selected regions in an array of electrodes hooked up to a camera always creates a batch of star-like images on a dark background. So how can one provide information about dark objects?

Given these basic facts, the question arises as to what the best approach is for converting visual images into electrical stimulation of implanted electrodes. We shall consider three possibilities. The first approach is to use what may be called a *sustained activator* with which the frequencies and/or current levels are set to be proportional to the light level within the cells of the camera unit, each of which connects to a specific electrode in the array. There are several reasons why such a system is undesirable: (1) Sustained activation does not appear to produce a constant star-like image; it has been shown that when electrical stimulation is prolonged, images in area V1 fade away (Schmidt et al., 1996). (2) Varying frequency and current levels produces changes in perceived contrast over only a relatively narrow range as has already been noted (Schiller et al., 2006). (3) Most neurons in area V1 do not respond in a sustained fashion to light; instead, they respond relatively briefly to *changes* in illumination. (4) To assure the longevity of implanted electrode arrays, duration of activation should be minimized. Taking these considerations into account a sustained activator system would seem most undesirable.

Another approach is to set up the prosthetic device as a *transient luminance difference* detector that activates the electrode array briefly whenever a change occurs within the individual cells of the camera that activates the individual electrodes. This arrangement would greatly reduce the extent to which electrodes are activated over time. A major problem that arises, however, is that a response is elicited both when a stimulus appears and when it disappears thereby interfering with the proper analysis of temporal events. An example demonstrating this appears in Display 1. This dynamic display, and subsequent ones, can be viewed on the journal's website. On the left of Display 1 the visual image is shown that consists of a horizontal and vertical line appearing repeatedly in succession. Shown on the right are the star-like images created by the activation of the electrodes in V1. Due to the fact that the *luminance difference detector* responds both when a stimulus appears and when it disappears, instead of seeing alternating horizontal and vertical lines, a stationary cross is perceived. In a more general way, this arrangement would do a poor job in handling motion.

A solution that overcomes these problems is to create a system in which each cell is turned into what we call an *intracell comparator* (Dobelle, 2000). This is a procedure used in various forms in a number of routines for visual processing in computer vision; it is often referred to as an edge detector system. The general idea is that each cell in the camera unit activates a corresponding electrode only when a change in illumination occurs *within* the elements of a cell. This arrangement is similar to how the majority of V1 cells respond as has been documented by recording from hundreds of sites in cat and monkey (Hubel and Wiesel, 1972, 1977, 2005; Schiller et al, 1976a&b), an example of which appears in Figure 10A. Here data are shown as obtained from an alert monkey trained to fixate. After mapping the receptive field of the multiple units from which we recorded, data were collected under three conditions: (1)

flashing on a small white spot for 500 milliseconds within the receptive field, (2) flashing on a small dark spot, and (3) flashing on a white spot that was larger than the receptive field. Each condition was presented 40 times. The neurons respond in a similar fashion to the small light and dark spots and do not respond at all to the large spot. Thus as a group these cells behave like *intracell comparators*.

Creating an *intracell comparator* is depicted in Figure 10B. In this arrangement each cell is hooked up to an electrode as had been described in Figures 7–10. When a change in illumination occurs that affects all elements *within* a cell equally, no response is elicited as depicted in b in Figure 10B since there is no differential activation *within* the cell. By contrast, when a luminance difference arises *within* the cell, as shown in d, f and h, a short-lasting activation results. The system has the option of providing different levels of activation based on magnitude of the contrast difference that arises within cells; the greater the contrast change within a cell, as shown in Figure 10d, f, and h, the higher the frequency or current produced. Since overall changes in perceived contrast as a function of frequency and current level fall within a narrow range as we had noted, only a limited number of levels could be used effectively. We believe that the *intracell comparator* system is one which is in consonance with the basic operational principles of neurons in V1 as shown on top of Figure 10A.

The *intracell comparator* system is single ended: A response is elicited to any sign of contrast change. Thus the response produced by black letters on a white background or white letters on a dark background produce similar activation. This arrangement is in consonance with the response characteristics of the majority of V1 cells. The response is made transient by having the activation occur only briefly, adjustable between perhaps 200 to 500 milliseconds to fit personal needs and situations.

Based on mimicking procedures it appears that the *intracell comparator* system works reasonably well. Display 2 shows this. Unlike in the previous display (Display 1), the alternating horizontal and vertical lines are reproduced quite faithfully. A more complex example of this appears in Displays 3 and 4. Both displays show the words FIAT LUX drifting across visual space, with Display 3 using the *temporal difference detector* scheme and Display 4 showing the *intracell comparator* system. The latter does a rather decent job in displaying the moving image.

The limitations of the *intracell comparator* system for processing images in two-dimensional space should be acknowledged, however. While outline forms with relatively well-defined edges or lines work quite well, smooth, extended surfaces are not visible, as indicated in Figure 10B. A dynamic example of this is shown in Display 5 in which the image created by a moving outline square is presented. The percept created corresponds nicely with the actual image. However, when a solid square is used, this is not the case, as shown in Display 6. Instead of a solid square, an outline square is perceived since the solid regions do not activate the cells except for the edges. One way to remedy this problem is to break up solid surfaces into grained surfaces. Display 7 shows the arrangement when a solid area in the display is changed to a crosshatched one. As a result of this, the image created appears more like a solid surface because the cross hatched regions keep activating the cells over which they course.

It should be instructive at this juncture to examine what kind of image is created using the *intracell comparator* system when an outline square is moved across the visual field using the butterfly array. The moving square arrangement is identical to the one shown in Display 5 for which the proportional array was used. Display 8 shows the images elicited using the equally spaced butterfly array depicted in Figure 9. The percept created only remotely resembles the actual display. What is seen instead is an object that changes shape and size rather than being constant. It would seem unlikely that even extensive training could overcome this problem and

enable a blind subject implanted with the butterfly array to perceive a square of a constant size moving across the visual field. Because of this, we believe that it is highly desirable to use proportional arrays that can provide unchanging images of objects as they move across the visual field successively activating different regions of area V1.

f. Procedures must be created to provide egocentric spatial cues

One central requirement for a visual prosthetic device is to provide information about where objects are in the visual scene relative to the body. Due to the noted fact that eye movements, accommodation and the vestibulo-ocular reflex are compromised in most blind patients, the incorporation of eye-movement signals in a prosthetic device is unlikely to solve the problem. Another, perhaps more promising approach, is to train blind subjects to minimize their eye movements and, in addition, to provide a *reference signal*. One such possible signal would be the differential activation of a region in area V1 that represents the center of the fovea. To make it differential, the electrical stimulation of this area could occur in brief bursts, thereby making it appear as a flickering phosphene. It would be activated, as needed, by the subject, perhaps by pushing a button. This independent signal, always denoting the same retinal location, could then be utilized to move the eyes to desired locations. For example, were a blind subject to place one of his fingers in front of the camera, the pulsing reference signal could be shifted by the movement of the eyes to be superimposed on the finger. The somatosensory system provides information about position of the finger relative to the body. As a result, the location of visual objects processed by the camera activating the implanted electrode array could be linked to the position of the finger, thereby providing cues about the location of objects in the visual scene relative to the body.

g. Arrangements must be made to optimize the processing of fine detail

There are two prime ways of optimizing fine detail. The first is to use a camera with a zoom. The second is to increase the number of electrodes implanted into the brain.

Using a zoom camera is extremely effective in improving fine detail as it has properties on a small scale similar to that of a microscope or telescope. A simple, obvious example is to prop up a book a few feet away and look at it through a camera. By zooming in the words will become readable. This procedure can therefore significantly enhance acuity when such a camera is hooked up to activate implanted electrode arrays as described in Figures 7–11.

To improve acuity by varying the density and numerosity of electrodes in implanted arrays two major limitations need to be noted. The first is that large arrays are likely to cause tissue damage either upon implantation or subsequently with the passage of time. The second is that the density of electrode placement is limited by the size of the phosphenes created. Very closely spaced electrodes would not create spatially distinguishable spots and therefore would be unlikely to improve acuity. When at a four degree eccentricity electrodes are placed 1.3 millimeters apart in monkey V1, we estimate that the phosphenes created have only a small degree of overlap as shown in Figures 6–8. Because more tissue is allocated for central vision, and because the phosphenes created become smaller when regions near the foveal representation are stimulated, in this region electrodes can be profitably placed closer to each other without the stimulation creating overlapping phosphenes.

So the question that arises then is how one can optimize resolution most effectively by proper electrode placing. Figure 11 shows two schemes. On the left, a proportional array is shown that is enhanced for the central 2 degrees of the visual field by doubling up on the number of electrodes in this region as indicated by the electrodes marked in blue in the bottom of Figure 11A. The electrode placements marked in red are the same as for the proportional arrays shown in Figure 7. This arrangement increases the number of electrodes from 128 per side to 224 per

side. The improvement in resolution in this central area is shown in 11B showing the activation produced by the word FIAT LUX that occupies just over a two degree region in the visual field viewed by the camera. To obtain similar resolution at eccentricities between 2 and 4 degrees from the center of the fovea with an equally spaced array, 2.85 times as many electrodes would be needed: 610 per side as depicted in 11C. In the proportional array further resolution could be gained by doubling up the electrodes for the central one degree of foveal vision which would produce spatially separate phosphenes within this region. Such placement would require 96 more electrodes but could double acuity within this region.

h. The prosthetic device must be able to provide information in three dimensions

As we had noted in section 2, one of the essential requirements for a prosthetic device is to be able to provide information about the relative location of objects in depth. Without such information locomotion in space would be next to impossible. We had also noted that a prosthetic device that relies on electrical stimulation should be capable of providing motion parallax cues for the extraction of depth information.

Here we provide two basic examples to indicate that the enhanced proportional array of 448 elements using the *intracell comparator* system is capable of providing a modicum of depth cues based on motion parallax. In Display 9 eight short vertical lines are displayed on the left which are rotated in steps to provide the impression of movement in depth. On the right is shown the manner in which visual impressions are presumed to be created by the electrical stimulation elicited by the camera viewing the display on the left. Although the image is degraded, most viewers can derive a sense of depth from the right display.

The second example is shown in Display 10. The image shown is a little bit more complex than the previous one. A drifting truncated pyramid is rocked back and forth across the visual field along its vertical axis. The sense of depth created with the enhanced proportional array reproduces the three-dimensional impression reasonably well.

i. Mimicking of the presumed effects must be carried out in sighted subjects to provide insights about the manner in which electrodes should be spaced and activated in an array

To study the visual impressions that might be created using a prosthetic device based on electrical stimulation of area V1, we have devised a system that converts the input from a camera mounted on the head into visual images viewed on a monitor in a bezel (as shown in Figure 12) that mimic the images created by electrical stimulation as described in Figures 7–9. The program is flexible and allows for the manipulation of various array configurations. Two possible arrangements are shown in Figure 13. On the left is the proportional array shown earlier, and on the right the proportional circular array. The camera views a display that consists of a large circle and a small square. The electrodes activated to produce the phosphenes appear in the top left and right sections. The resultant images created appear on the bottom.

With this arrangement we can proceed to determine how effective a prosthetic device might be in a real life situation when free movement is allowed and when the visual scene is dynamic. Subjects are free to move their heads about so they can optimize the process of deciphering the visual scene. To learn about how they move their heads, a laser is attached to the camera; consequently the location of the laser spot in the visual scene can be traced enabling the experimenter to determine what strategies subjects use to decipher the visual scene (e.g. Dobelle, 2000). Furthermore, subjects can be provided with a controller for the camera zoom which enables them to optimize the resolution of the images viewed.

Using this mimicking system one can address the following central questions: (1) How well can the system process stationary images? (2) How can head movement improve processing?

(3) How can acuity be optimized by manipulating the zoom feature on the camera and by mimicking different array configurations? (4) How can moving objects be analyzed best? (5) How can depth perception be realized? (6) How can one effectively minimize the amount of activation of brain tissue yet effectively process visual displays? Our preliminary tests show that the percepts created using the mimicking device depicted in Figure 12 set to convert the camera image with the *temporal intracell comparator* yields images similar to those shown in Displays 4–10.

7. A brief description of one experimental session

Here is an excerpt from our procedure with one volunteer:

The helmet with the camera and the bezel with the small monitor was placed on his head. We used the proportional square array described in Figure 7 and the *temporal intracell comparator*. The subject was instructed to direct his head toward the wall on which was pasted the famous statement made by John F. Kennedy at his inaugural address in 1961. The words appeared in a single line and the subject was asked to first activate the camera zoom to optimize resolution and to then scan the display by slowly moving his head from left to right:

ASK NOT WHAT YOUR COUNTRY CAN DO FOR YOU – ASK WHAT YOU CAN DO FOR YOUR COUNTRY

The image created looked similar to what is shown in Display 11. The subject recited the words clearly. As he spoke, the experimenter detected a slight accent, so he asked where he was from. “I am Italian,” he said. “From Tuscany,” he added.

“All right, in that case I would like to show you one more display,” said the experimenter. “Just keep your head still and I will flash on a figure. You tell me what you see.” The experimenter flashed on a circle using the proportional circular array (Figures 8 and 12) which then appeared on the monitor; the image created looked like the one shown in Display 12.

“Okay, what do you see?” the experimenter inquired.

“I see a circle. The round O of Giotto,” the subject added.

DYNAMIC DISPLAYS

Display 1

The responses elicited in a proportional square array to an alternating horizontal and vertical line using the *transient luminance difference* detector system. The visual stimulus appears on the left and the activation producing the visual impression created by the electrical stimulation is shown on the right. Each line is shown for 180 ms. Activation duration is set to last 90 milliseconds. Due to a response elicited both when the stimulus appears and is terminated, a persistent cross is seen instead of alternating lines.

Display 2

The responses elicited in a proportional square square array to an alternating horizontal and vertical line using the *temporal intracell comparator* system. The visual stimulus appears on the left and the activation producing the visual impression created by the electrical stimulation is shown on the right. Each line is shown for 180 ms. Activation duration is set to last 90 milliseconds. The alternating lines are reproduced quite well.

Display 3

The responses elicited in a proportional square array to the drifting words FIAT LUX using the *temporal difference detector* system. The visual stimulus appears on the left and the activation producing the visual impression created by the electrical stimulation is shown on the right. The letters move across the visual field in 90 millisecond steps. Activation duration is set to 90 milliseconds.

Display 4

The responses elicited in a proportional square array to the drifting words FIAT LUX using the *temporal intracell comparator* system. The visual stimulus appears on the left and the activation producing the visual impression created by the electrical stimulation is shown on the right. The letters move across the visual field in 90 millisecond steps. Activation duration is set to 90 milliseconds. The reproduction using this scheme is superior to the one shown is superior to the one shown in Display 3 that uses the *temporal difference detector* system.

Display 5

The responses elicited in a proportional array to a drifting outline square using the *temporal intracell comparator* system. The visual stimulus appears on the left and the activation producing the visual impression created by the electrical stimulation is shown on the right. The display moves across the visual field in 90 millisecond steps. Activation duration is set to 90 milliseconds. The square is reproduced quite faithfully.

Display 6

The responses elicited in a proportional square array to a drifting solid square using the *temporal intracell comparator* system. The display moves across the visual field in 90 milliseconds steps. Activation duration is set to 90 milliseconds. This system produces an outline square instead of a solid square.

Display 7

The responses elicited in a proportional square array to a drifting cross-hatched square using the *temporal intracell comparator* system. The display moves across the visual field in 90 millisecond steps. Activation duration is set to 90 milliseconds. The cross hatching in the drifting visual display now produces what is pretty much a solid square.

Display 8

The responses elicited in an equally spaced (butterfly) array to a drifting outline square using the *temporal intracell comparator* system. The display moves across the visual field in 90 millisecond steps. Activation duration is set to 90 milliseconds. Instead of a square moving in space an object is seen that undergoes notable changes in shape from which it would be difficult to infer the movement of a single, invariant moving object.

Display 9

The responses elicited in a proportional square array to a group of rotating lines using *temporal intracell comparator* system. The impression of three dimensions is reasonably well preserved.

Display 10

The responses elicited in a proportional square array to a rocking pyramid using the *temporal intracell comparator* system. The impression of three dimensions is reasonably well preserved.

Display 11

The image created when the text of John F. Kennedy's famous remark, pasted on the wall, was scanned by a subject. The camera was set to use the *temporal inrarcell comparator system* with the proportional square array as shown in Figure 7.

Display 12

The image created when a circle is flashed on after central fixation using the camera set to use the proportional circular array system.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

The authors wish to thank Christina E. Carvey, Warren M. Slocum, Geoffrey Kendall and David Feeney for their contributions in realizing this manuscript.

References

- Barlow HB. The Ferrier Lecture, 1980. Critical limiting factors in the design of the eye and visual cortex. *Proc R Soc Lond B Biol Sci* 1981;212:1–34. [PubMed: 6115386]
- Barlow HB, Blakemore C, Pettigrew JD. The neural mechanism of binocular depth discrimination. *J Physiol* 1967;193:327–342. [PubMed: 6065881]
- Barlow HB, Levick WR. Changes in the maintained discharge with adaptation level in the cat retina. *J Physiol* 1969;202:699–718. [PubMed: 5789945]
- Bartels M. Beobachtungen an Wirbeltieren und Menschen über unwillkürliche Augenbewegungen bei Störungen des Sehens. 2. Beobachtungen an Menschen. *Klin Monatsbl Augenheilkd* 1928;80:145–176.
- Bartlett JR, DeYoe EA, Doty RW, Lee BB, Lewine JD, Negrao N, Overman WH Jr. Psychophysics of electrical stimulation of striate cortex in macaques. *J Neurophysiol* 2005;94:3430–3442. [PubMed: 16079195]
- Bartlett JR, Doty RW Sr. Response of units in striate cortex of squirrel monkeys to visual and electrical stimuli. *J Neurophysiol* 1974;37:621–641. [PubMed: 4209701]
- Bloomfield SA, Dowling JE. Roles of aspartate and glutamate in synaptic transmission in rabbit retina. I. Outer plexiform layer. *J Neurophysiol* 1985;53:699–713. [PubMed: 2858516]
- Bradley DC, Troyk PR, Berg JA, Bak M, Cogan S, Erickson R, Kufta C, Mascaro M, McCreery D, Schmidt EM, et al. Visuotopic mapping through a multichannel stimulating implant in primate V1. *J Neurophysiol* 2005;93:1659–1670. [PubMed: 15342724]
- Brindley GS, Lewin WS. The sensations produced by electrical stimulation of the visual cortex. *J Physiol* 1968;196:479–493. [PubMed: 4871047]
- Broos BPJ. The 'O' of Rembrandt. *Simiolus: Netherlands Quarterly for the History of Art* 1971;4:150–184.
- Casagrande VA, Boyd JD. The neural architecture of binocular vision. *Eye* 1996;10(Pt 2):153–160. [PubMed: 8776442]
- Chatterjee S, Callaway EM. Parallel colour-opponent pathways to primary visual cortex. *Nature* 2003;426:668–671. [PubMed: 14668866]
- Chino YM, Kaas JH, Smith EL 3rd, Langston AL, Cheng H. Rapid reorganization of cortical maps in adult cats following restricted deafferentation in retina. *Vision Res* 1992;32:789–796. [PubMed: 1604848]
- Clark, G. Cochlear implants fundamentals and applications. New York: Springer; 2003.

- Clark GM. The multiple-channel cochlear implant: the interface between sound and the central nervous system for hearing, speech, and language in deaf people—a personal perspective. *Philos Trans R Soc Lond B Biol Sci* 2006;361:791–810. [PubMed: 16627295]
- Cohen ED. Prosthetic interfaces with the visual system: biological issues. *J Neural Eng* 2007;4:R14–31. [PubMed: 17409473]
- Cowey A, Walsh V. Magnetically induced phosphenes in sighted, blind and blindsighted observers. *Neuroreport* 2000;11:3269–3273. [PubMed: 11043562]
- Darian-Smith C, Gilbert CD. Topographic reorganization in the striate cortex of the adult cat and monkey is cortically mediated. *J Neurosci* 1995;15:1631–1647. [PubMed: 7891124]
- Demer JL, Echelman DA, Robinson DA. Effects of electrical stimulation and reversible lesions of the olivocerebellar pathway on Purkinje cell activity in the flocculus of the cat. *Brain Res* 1985;346:22–31. [PubMed: 3876867]
- Dobelle WH. Artificial vision for the blind by connecting a television camera to the visual cortex. *Asaio J* 2000;46:3–9. [PubMed: 10667705]
- Dobelle WH, Mladejovsky MG, Evans JR, Roberts TS, Girvin JP. “Braille” reading by a blind volunteer by visual cortex stimulation. *Nature* 1976;259:111–112. [PubMed: 1246346]
- Dobelle WH, Mladejovsky MG, Girvin JP. Artificial vision for the blind: electrical stimulation of visual cortex offers hope for a functional prosthesis. *Science* 1974;183:440–444. [PubMed: 4808973]
- Felleman DJ, Van Essen DC. Distributed hierarchical processing in the primate cerebral cortex. *Cereb Cortex* 1991;1:1–47. [PubMed: 1822724]
- Fitzpatrick D, Lund JS, Blasdel GG. Intrinsic connections of macaque striate cortex: afferent and efferent connections of lamina 4C. *J Neurosci* 1985;5:3329–3349. [PubMed: 3001242]
- Freeman RD, Ohzawa I. On the neurophysiological organization of binocular vision. *Vision Res* 1990;30:1661–1676. [PubMed: 2288082]
- Gilbert CD, Hirsch JA, Wiesel TN. Lateral interactions in visual cortex. *Cold Spring Harb Symp Quant Biol* 1990;55:663–677. [PubMed: 2132846]
- Gilbert CD, Wiesel TN. Receptive field dynamics in adult primary visual cortex. *Nature* 1992;356:150–152. [PubMed: 1545866]
- Grèusser, O-J. Light not illuminating the world: phosphenes and related phenomena. In: Grèusser, O-J.; Landis, T., editors. *Visual agnosias and other disturbances of visual perception and cognition, Vision and visual dysfunction*. Vol. 12. Boca Raton: CRC Press; 1991. p. 158–178.
- Hall EC, Ciuffreda KJ. Fixational ocular motor control is plastic despite visual deprivation. *Vis Neurosci* 2002;19:475–481. [PubMed: 12511080]
- Hartline HK. The responses of single optic nerve fibers of the vertebrate eye to illumination of the retina. *Am J Physiol* 1938;121:400–415.
- Heinen SJ, Skavenski AA. Recovery of visual responses in foveal V1 neurons following bilateral foveal lesions in adult monkey. *Exp Brain Res* 1991;83:670–674. [PubMed: 2026207]
- Hendry SH. Delayed reduction in GABA and GAD immunoreactivity of neurons in the adult monkey dorsal lateral geniculate nucleus following monocular deprivation or enucleation. *Exp Brain Res* 1991;86:47–59. [PubMed: 1756798]
- Hendry SH, Yoshioka T. A neurochemically distinct third channel in the macaque dorsal lateral geniculate nucleus. *Science* 1994;264:575–577. [PubMed: 8160015]
- Howard, IP. Seeing in depth, Vol Volume 1: Basic Mechanisms. Toronto: I. Porteous; 2002.
- Hubel DH, Wiesel TN. Laminar and columnar distribution of geniculate-cortical fibers in the macaque monkey. *J Comp Neurol* 1972;146:421–450. [PubMed: 4117368]
- Hubel DH, Wiesel TN. Ferrier lecture. Functional architecture of macaque monkey visual cortex. *Proc R Soc Lond B Biol Sci* 1977;198:1–59. [PubMed: 20635]
- Hubel, DH.; Wiesel, TN. *Brain and visual perception: the story of a 25-year collaboration*. New York, N.Y: Oxford University Press; 2005.
- Humayun MS, de Juan E Jr, Weiland JD, Dagnelie G, Katona S, Greenberg R, Suzuki S. Pattern electrical stimulation of the human retina. *Vision Res* 1999;39:2569–2576. [PubMed: 10396625]

- Humayun MS, Weiland JD, Fujii GY, Greenberg R, Williamson R, Little J, Mech B, Cimarusti V, Van Boemel G, Dagnelie G, de Juan E. Visual perception in a blind subject with a chronic microelectronic retinal prosthesis. *Vision Res* 2003;43:2573–2581. [PubMed: 13129543]
- Jones GM. Plasticity in the adult vestibulo-ocular reflex arc. *Philos Trans R Soc Lond B Biol Sci* 1977;278:319–334. [PubMed: 19786]
- Jones GM, Berthoz A, Segal B. Adaptive modification of the vestibulo-ocular reflex by mental effort in darkness. *Exp Brain Res* 1984;56:149–153. [PubMed: 6332032]
- Kaas JH, Krubitzer LA, Chino YM, Langston AL, Polley EH, Blair N. Reorganization of retinotopic cortical maps in adult mammals after lesions of the retina. *Science* 1990;248:229–231. [PubMed: 2326637]
- Karten JH, Fite KV, Brecha N. Specific projection of displaced retinal ganglion cells upon the accessory optic system in the pigeon (*Columba livia*). *Proc Natl Acad Sci U S A* 1977;74:1753–1756. [PubMed: 266216]
- Kayama Y, Riso RR, Bartlett JR, Doty RW. Luxotonic responses of units in macaque striate cortex. *J Neurophysiol* 1979;42:1495–1517. [PubMed: 115968]
- Klatzky, RL. Allocentric and egocentric spatial representations: definitions, distinctions, and interconnections. In: Freksa, C.; Habel, C.; Wender, KF., editors. *Spatial cognition: an interdisciplinary approach to representing and processing spatial knowledge*. Berlin ; New York: Springer; 1998. p. 1-17.
- Kömpf D, Piper HF. Eye movements and vestibulo-ocular reflex in the blind. *J Neurol* 1987;234:337–341. [PubMed: 3612205]
- Kuffler SW. Discharge patterns and functional organization of mammalian retina. *J Neurophysiol* 1953;16:37–68. [PubMed: 13035466]
- Legge GE, Pelli DG, Rubin GS, Schleske MM. Psychophysics of reading--I. Normal vision. *Vision Res* 1985;25:239–252. [PubMed: 4013091]
- Leigh RJ, Zee DS. Eye movements of the blind. *Invest Ophthalmol Vis Sci* 1980;19:328–331. [PubMed: 6965668]
- Leonard, R. *Statistics on Vision Impairment: A Resource Manual*. Arlene R. Gordon Research Institute of Lighthouse International; 2002. p. 1-49.
- LeVay S, Hubel DH, Wiesel TN. The pattern of ocular dominance columns in macaque visual cortex revealed by a reduced silver stain. *J Comp Neurol* 1975;159:559–576. [PubMed: 1092736]
- Malpeli JG, Baker FH. The representation of the visual field in the lateral geniculate nucleus of *Macaca mulatta*. *J Comp Neurol* 1975;161:569–594. [PubMed: 1133232]
- Marg E, Dierssen G. Reported visual percepts from stimulation of the human brain with microelectrodes during therapeutic surgery. *Confin Neurol* 1965;26:57–75. [PubMed: 5325250]
- Martin PR, White AJ, Goodchild AK, Wilder HD, Sefton AE. Evidence that blue-on cells are part of the third geniculocortical pathway in primates. *Eur J Neurosci* 1997;9:1536–1541. [PubMed: 9240412]
- Matthews MR, Cowan WM, Powell TP. Transneuronal cell degeneration in the lateral geniculate nucleus of the macaque monkey. *J Anat* 1960;94:145–169. [PubMed: 17105113]
- Mellor, CM. *Louis Braille: a touch of genius*. Boston, Mass: National Braille Press; 2006.
- Mize RR, Luo Q, Tigges M. Monocular enucleation reduces immunoreactivity to the calcium-binding protein calbindin 28 kD in the rhesus monkey lateral geniculate nucleus. *Vis Neurosci* 1992;9:471–482. [PubMed: 1450101]
- O’Kusky J, Colonnier M. A laminar analysis of the number of neurons, glia, and synapses in the adult cortex (area 17) of adult macaque monkeys. *J Comp Neurol* 1982;210:278–290. [PubMed: 7142443]
- Perry VH, Cowey A. The ganglion cell and cone distributions in the monkey’s retina: implications for central magnification factors. *Vision Res* 1985;25:1795–1810. [PubMed: 3832605]
- Peters, A. Number of neurons and synapses in primary visual cortex. In: Jones, EG.; Peters, A., editors. *Cerebral cortex*. Vol. 6. New York: Plenum Press; 1987. p. 267-294.
- Peters, A. The organization of the primary visual cortex. In: Peters, A.; Rockland, KS., editors. *Cerebral cortex*. Vol. 10. New York: Plenum Press; 1994. p. 1-35.
- Pezaris, JS.; Reid, RC. The color of percepts evoked by microstimulation in macaque LGN. Program No. 13.5. Neuroscience Meeting Planner; Atlanta, GA: Society for Neuroscience; 2006. Online

- Pezaris JS, Reid RC. Demonstration of artificial visual percepts generated through thalamic microstimulation. *Proc Natl Acad Sci U S A* 2007;104:7670–7675. [PubMed: 17452646]
- Poggio GF, Fischer B. Binocular interaction and depth sensitivity in striate and prestriate cortex of behaving rhesus monkey. *J Neurophysiol* 1977;40:1392–1405. [PubMed: 411898]
- Pritchard RM. Stabilized images on the retina. *Sci Am* 1961;204:72–78. [PubMed: 13738093]
- Richards W. The fortification illusions of migraines. *Sci Am* 1971;224:88–96. [PubMed: 5552581]
- Rizzo JF 3rd, Wyatt J, Loewenstein J, Kelly S, Shire D. Perceptual efficacy of electrical stimulation of human retina with a microelectrode array during short-term surgical trials. *Invest Ophthalmol Vis Sci* 2003a;44:5362–5369. [PubMed: 14638739]
- Rizzo JF, Jensen RJ, Loewenstein J, WJL. Unexpectedly small percepts evoked by epi-retinal electrical stimulation in blind humans. *Invest Ophthalmol Vis Sci* 2003b;44:4207.
- Robinson DA. Eye movements evoked by collicular stimulation in the alert monkey. *Vision Res* 1972;12:1795–1808. [PubMed: 4627952]
- Rockel AJ, Hiorns RW, Powell TP. The basic uniformity in structure of the neocortex. *Brain* 1980;103:221–244. [PubMed: 6772266]
- Rodieck, RW. The vertebrate retina; principles of structure and function. San Francisco: California, Freeman; 1973.
- Roe AW, Parker AJ, Born RT, DeAngelis GC. Disparity channels in early vision. *J Neurosci* 2007;27:11820–11831. [PubMed: 17978018]
- Rushton, DN.; Brindley, GS. Short- and long-term stability of cortical electrical phosphenes. In: Rose, FC., editor. *Physiological aspects of clinical neurology*. Oxford: Blackwell Scientific Publications; 1977. p. 123-153.
- Santos A, Humayun MS, de Juan E Jr, Greenburg RJ, Marsh MJ, Klock IB, Milam AH. Preservation of the inner retina in retinitis pigmentosa. A morphometric analysis. *Arch Ophthalmol* 1997;115:511–515. [PubMed: 9109761]
- Schein SJ. Anatomy of macaque fovea and spatial densities of neurons in foveal representation. *J Comp Neurol* 1988;269:479–505. [PubMed: 3372725]
- Schiller, PH. The ON and OFF channels of the mammalian visual system. In: Osborne, NN.; Chader, GJ., editors. *Progress in Retinal and Eye Research*. Oxford: England, Pergamon Press; 1996. p. 173-195.
- Schiller PH, Finlay BL, Volman SF. Quantitative studies of single-cell properties in monkey striate cortex. I. Spatiotemporal organization of receptive fields. *J Neurophysiol* 1976a;39:1288–1319. [PubMed: 825621]
- Schiller PH, Finlay BL, Volman SF. Quantitative studies of single-cell properties in monkey striate cortex. II. Orientation specificity and ocular dominance. *J Neurophysiol* 1976b;39:1320–1333. [PubMed: 825622]
- Schiller PH, Logothetis NK. The color-opponent and broad-band channels of the primate visual system. *Trends Neurosci* 1990;13:392–398. [PubMed: 1700509]
- Schiller PH, Logothetis NK, Charles ER. Functions of the colour-opponent and broad-band channels of the visual system. *Nature* 1990a;343:68–70. [PubMed: 2296292]
- Schiller PH, Logothetis NK, Charles ER. Role of the color-opponent and broad-band channels in vision. *Vis Neurosci* 1990b;5:321–346. [PubMed: 2265148]
- Schiller PH, Malpeli JG. Properties and tectal projections of monkey retinal ganglion cells. *J Neurophysiol* 1977;40:428–445. [PubMed: 403252]
- Schiller PH, Stryker M. Single-unit recording and stimulation in superior colliculus of the alert rhesus monkey. *J Neurophysiol* 1972;35:915–924. [PubMed: 4631839]
- Schiller PH, Tehovnik EJ. Look and see: how the brain moves your eyes about. *Prog Brain Res* 2001;134:127–142. [PubMed: 11702539]
- Schiller PH, Tehovnik EJ, Weiner VS. Preliminary studies examining the feasibility of a visual prosthetic device. 2. The laminar specificity of electrical stimulation in monkey V1 and the visual percepts created. *J Vis* 2006;6:404a.

- Schmidt EM, Bak MJ, Hambrecht FT, Kufta CV, O'Rourke DK, Vallabhanath P. Feasibility of a visual prosthesis for the blind based on intracortical microstimulation of the visual cortex. *Brain* 1996;119 (Pt 2):507–522. [PubMed: 8800945]
- Schultze, M. The Eye. The Retina. In: Stricker, S., editor. *Manual of Human and Comparative Histology*. Vol. 3. London: The New Sydenham Society; 1873. p. 218–289.
- Schwartz, EL. Computational studies of the spatial architecture of primate visual cortex: columns, maps, and protomaps. In: Peters, A.; Rockland, KS., editors. *Cerebral cortex*. Vol. 10. New York: Plenum Press; 1994. p. 359–411.
- Sherman KR, Keller EL. Vestibulo-ocular reflexes of adventitiously and congenitally blind adults. *Invest Ophthalmol Vis Sci* 1986;27:1154–1159. [PubMed: 3487529]
- Simpson, JR.; Soodak, RE.; Hess, R. I. B. R. Organization. The accessory optic system and its relation to the vestibulocerebellum. In: Granit, R.; Pompeiano, O., editors. *Progress in brain research*, Vol. 50: Reflex control of posture and movement. Amsterdam: Elsevier; 1979.
- Sloper JJ, Headon MP, Powell TP. Changes in the size of cells in the monocular segment of the primate lateral geniculate nucleus during normal development and following visual deprivation. *Brain Res* 1987a;428:267–276. [PubMed: 3828834]
- Sloper JJ, Headon MP, Powell TP. Effects of enucleation at different ages on the sizes of neurons in the lateral geniculate nucleus of infant and adult monkeys. *Brain Res* 1987b;428:259–265. [PubMed: 3828833]
- Smirnakis SM, Brewer AA, Schmid MC, Toliás AS, Schuz A, Augath M, Inhoffen W, Wandell BA, Logothetis NK. Lack of long-term cortical reorganization after macaque retinal lesions. *Nature* 2005;435:300–307. [PubMed: 15902248]
- Stone JL, Barlow WE, Humayun MS, de Juan E Jr, Milam AH. Morphometric analysis of macular photoreceptors and ganglion cells in retinas with retinitis pigmentosa. *Arch Ophthalmol* 1992;110:1634–1639. [PubMed: 1444925]
- Tehovnik EJ. Electrical stimulation of neural tissue to evoke behavioral responses. *J Neurosci Methods* 1996;65:1–17. [PubMed: 8815302]
- Tehovnik EJ, Slocum WM. Phosphene induction by microstimulation of macaque V1. *Brain Res Rev* 2007a;53:337–343. [PubMed: 17173976]
- Tehovnik EJ, Slocum WM. What delay fields tell us about striate cortex. *J Neurophysiol* 2007b;98:559–576. [PubMed: 17567774]
- Troyk P, Bak M, Berg J, Bradley D, Cogan S, Erickson R, Kufta C, McCreery D, Schmidt E, Towle V. A model for intracortical visual prosthesis research. *Artif Organs* 2003;27:1005–1015. [PubMed: 14616519]
- Wagenaar, DA. Cortical stimulation for the evocation of visual perception. In: Andersen, R., editor. *Term paper for CNS247: Cerebral cortex*. Caltech; 2004. Retrieved on August 21, 2007 from <http://www.its.caltech.edu/~pinelab/wagenaar/writings.html>
- Wässle H, Boycott BB. Functional architecture of the mammalian retina. *Physiol Rev* 1991;71:447–480. [PubMed: 2006220]
- Weiland JD, Humayun MS. Past, present, and future of artificial vision. *Artif Organs* 2003;27:961–962. [PubMed: 14616514]
- Werblin FS, Dowling JE. Organization of the retina of the mudpuppy, *Necturus maculosus*. II. Intracellular recording. *J Neurophysiol* 1969;32:339–355. [PubMed: 4306897]
- Wiesel TN, Hubel DH. Spatial and chromatic interactions in the lateral geniculate body of the rhesus monkey. *J Neurophysiol* 1966;29:1115–1156. [PubMed: 4961644]

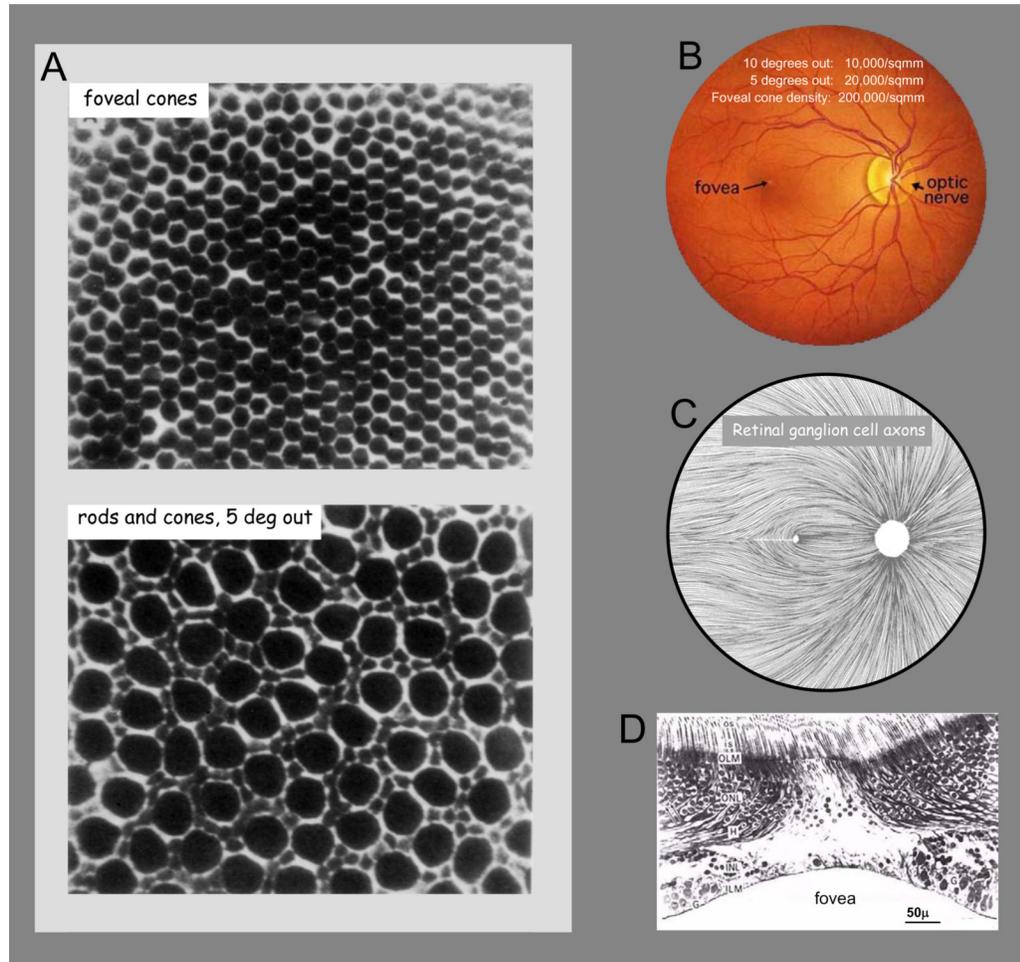


Figure 1.

The retina: A. Head-on view of the rods and cones in the fovea and at an eccentricity of 5 degrees. B. Head-on view of the retina showing the fovea, the optic disk and the blood vessels. C. Head-on view of the retina showing the course of the axons to the optic disk. D. Cross section of the retina at the foveal pit demonstrating that this region is free of retinal cells other than the photoreceptors; this is achieved by having cell processes course away from this central area.

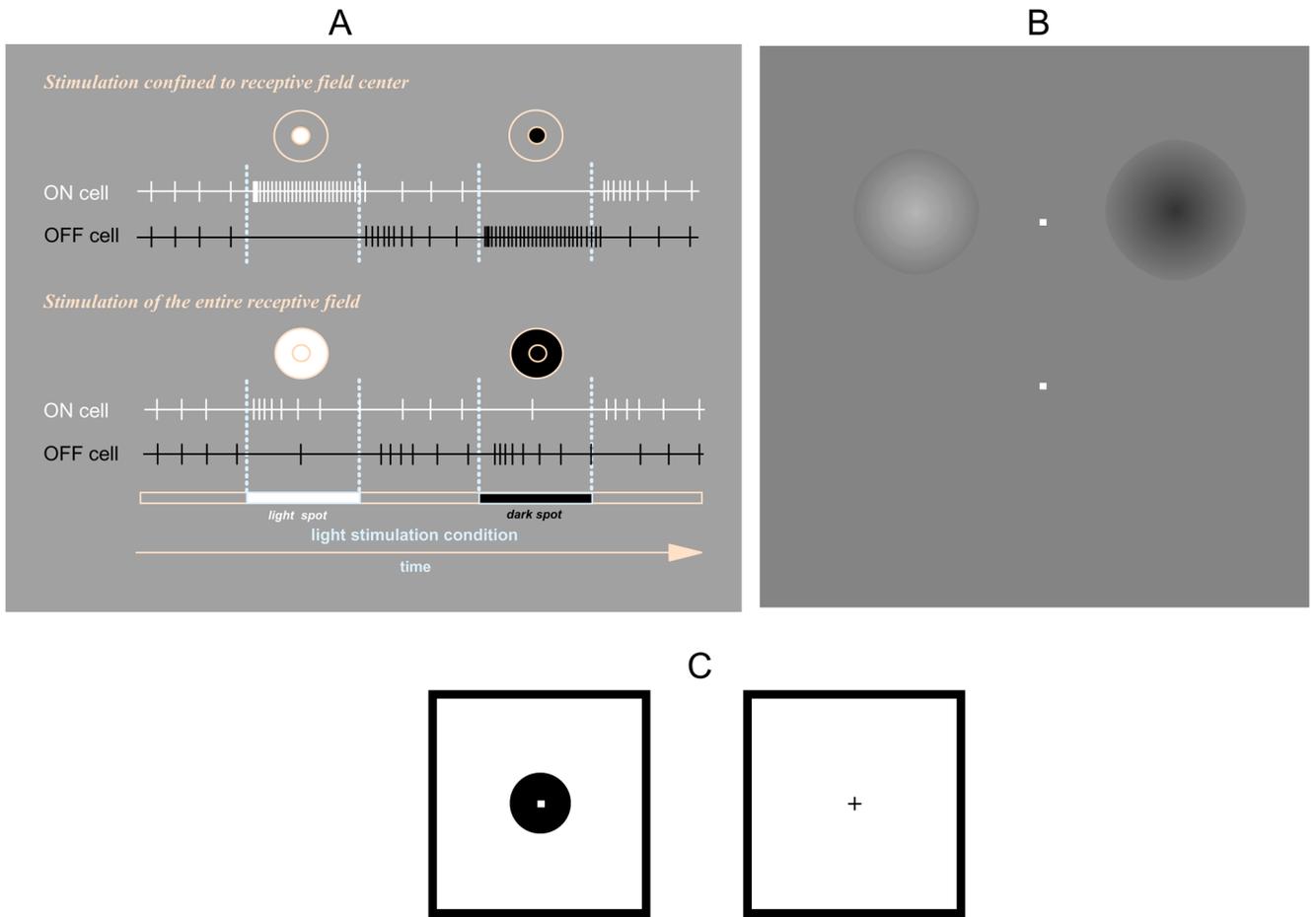


Figure 2.

A: Schematic of the responses of ON and OFF-center ganglion cells to spots presented in the center of their receptive field and large spots that impinge on both the center and the surround of their receptive fields. The vertical lines represent the action potentials created by the stimulation. Center-surround organization in these ganglion cells is antagonistic as a result of which a more vigorous response is elicited when a small spot is presented than a large one. ON-center ganglion cells are excited by light increment (the white spot on the gray background) whereas OFF-center cells are excited by light decrement (black spot on the gray background). B. Demonstration of the effects of adaptation. Fixating on the little white square in the center between the two gaussian disks for 10–20 seconds will result in the disappearance of these disks due to the adaptation process that takes place in the retina. At this point, when one shifts the center of gaze to the lower little white square, two negative afterimages will appear as a result of the photons from the homogeneous background impinging on more and less sensitive areas in the retina at the adapted locations. C. The scaling of afterimages. Fixate on the white dot in the center of the black disk in the left for 20–30 seconds, then shift your gaze to the black cross in the center of the right. Once the afterimage appears move the sheet toward and away from you. The afterimage will scale in size.

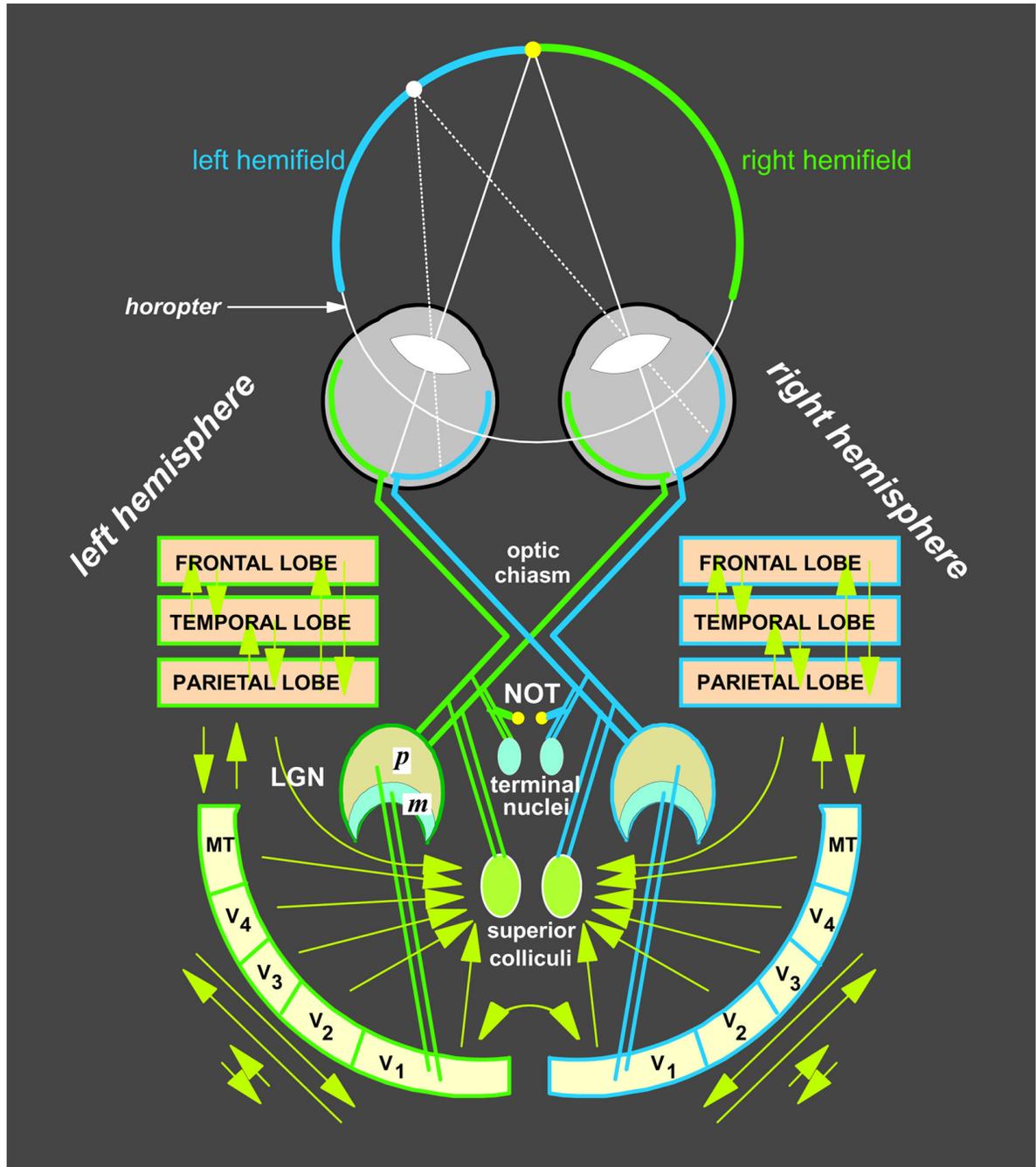


Figure 3.

The basic wiring diagram of the visual system. Ganglion cells from the nasal hemiretinae cross over at the optic chiasm whereas those from the temporal hemiretinae project ipsilaterally. As a result, punctate images in the visual field activate corresponding points in the visual system when they are presented along the horopter where an image activates corresponding points on the retinal surface. Images from the left visual hemifield (in blue) project to the right hemisphere and images from the right visual hemifield (green) project to the left hemisphere. The major projection sites from the retina are the lateral geniculate nucleus, the superior colliculus, the terminal nuclei. In the cortex there are numerous visual area (shown are V1, V2,

V3, V4, and MT). These areas send both feed forward and feed back connections to many areas of the brain. Images appear outside the horopter impinge on retinal non-corresponding points.

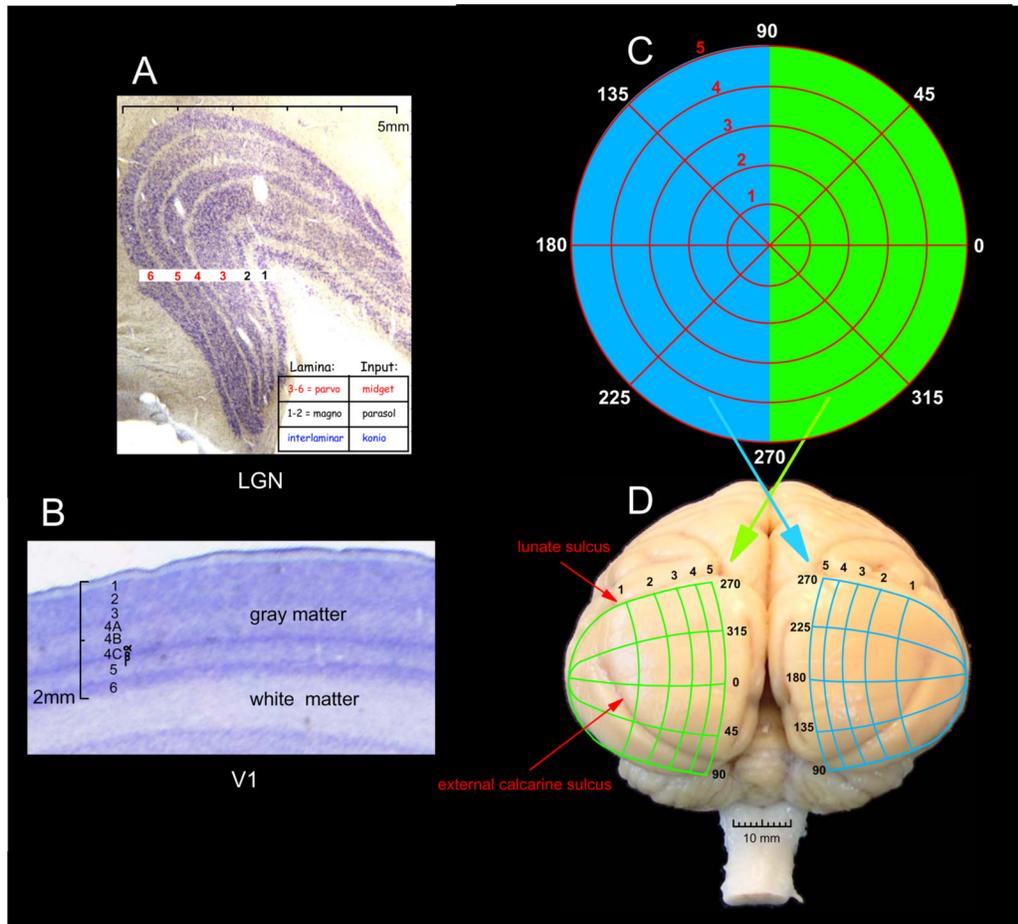


Figure 4.

Layout of the lateral geniculate nucleus and primary visual cortex (V1). A: A Nissyl-stained cross section of the lateral geniculate nucleus which, for the central 17 degrees of the visual field representation, has six layers. The top four layers (3–6) receive input from the retinal midget cells. The bottom two layers (1–2) receive input from the parasol cells. The intralaminar layers receive input mostly from the retinal koniocellular. B: A Nissyl-stained cross section of area V1. The thickness of gray matter and density of neurons is quite constant throughout this area. The layers in the gray matter are designated. The prime input of the midget and parasol cells from the LGN goes to layers 4c α and 4c β . The inputs from the koniocellular cells terminate in layers 1 and 2. C: Layout of the central five degrees of the visual field. The left hemifield (blue) projects to the right hemisphere and the right hemifield (green) projects to the left visual field. D: A rear view of the monkey brain showing the central 6–8 degrees of the visual field layout. This region in the monkey is lissencephalic except for the external calcarine sulcus, which is quite shallow. The visual field is laid out in a topographic fashion with much more area allocated for central than peripheral representation. The visual field is laid out upside down in the cortex with the upper part of the visual field in the lower region of V1 and the lower visual field in the upper portion of V1 using the conformal mapping scheme of Schwartz (1994) for the macaque monkey.

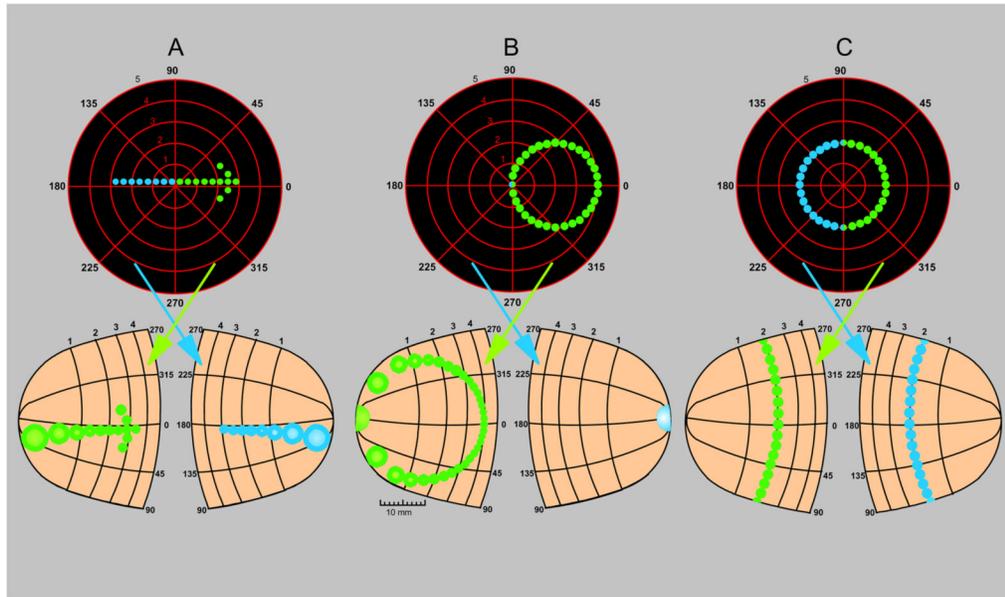


Figure 5.

The basic manner in which images activate various regions of area V1 in the monkey. Dotted figures are presented in the visual field in accordance with the fact that electrical stimulation of area V1 produces star-like images. A: A dotted arrow is placed in the visual field centered on the vertical meridian. The areas activated in V1 are shown below. The tail of the arrow projects into the right and the head of the arrow to the left hemisphere. Due to the magnification factor that results in more space allocated per unit area for central than for peripheral vision, the size of the area activated by each dot is progressively larger the closer the dots are to the foveal representation. Due to center/surround antagonism in retinal and LGN cells and due to the greater responses elicited to edges in the cortex, the dots drive neurons more vigorously at the outer periphery of each dot than in the center, which is depicted by the shading of the dots on the cortical surface. B: The dotted circle placed in the right visual hemifield activates the marked regions in the left hemifield; the dot in the center of the fovea activates the foveal representation in both hemispheres. The activation in the left hemifield forms a pretty good circle but the size of each dot representation changes as a function of eccentricity. C: The same circle is presented in the visual field centered on the vertical meridian. The activation in the cortex forms two crescents yet what we perceive is a perfect circle. The size of the areas activated is constant as these locations are equidistant from the fovea.

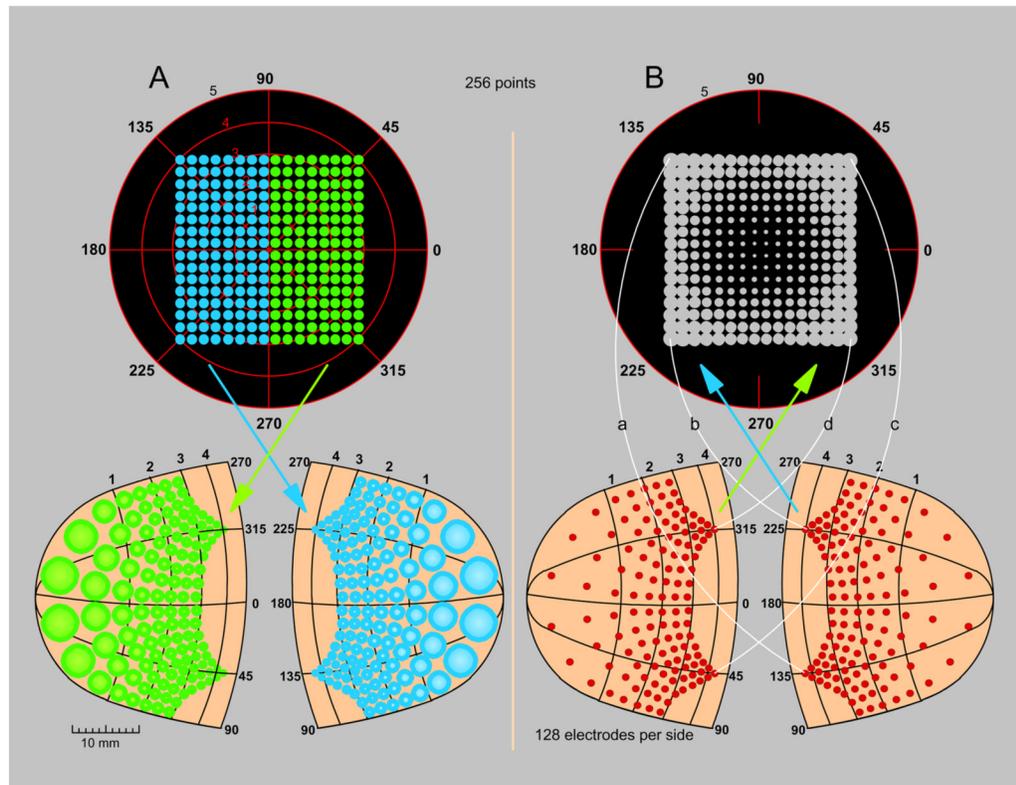


Figure 6.

A: An array of 256 dots arranged in the shape of a square and the corresponding brain regions activated in area V1. B: An array of 256 electrodes placed proportionally on the cortical surface taking magnification factor into account, with 128 in each hemisphere. Electrical stimulation under these conditions presumably activates an array or star-like images whose size increases with increasing eccentricity. The white lines show schematically the correspondence of points in the visual field and with the electrodes in the cortex.

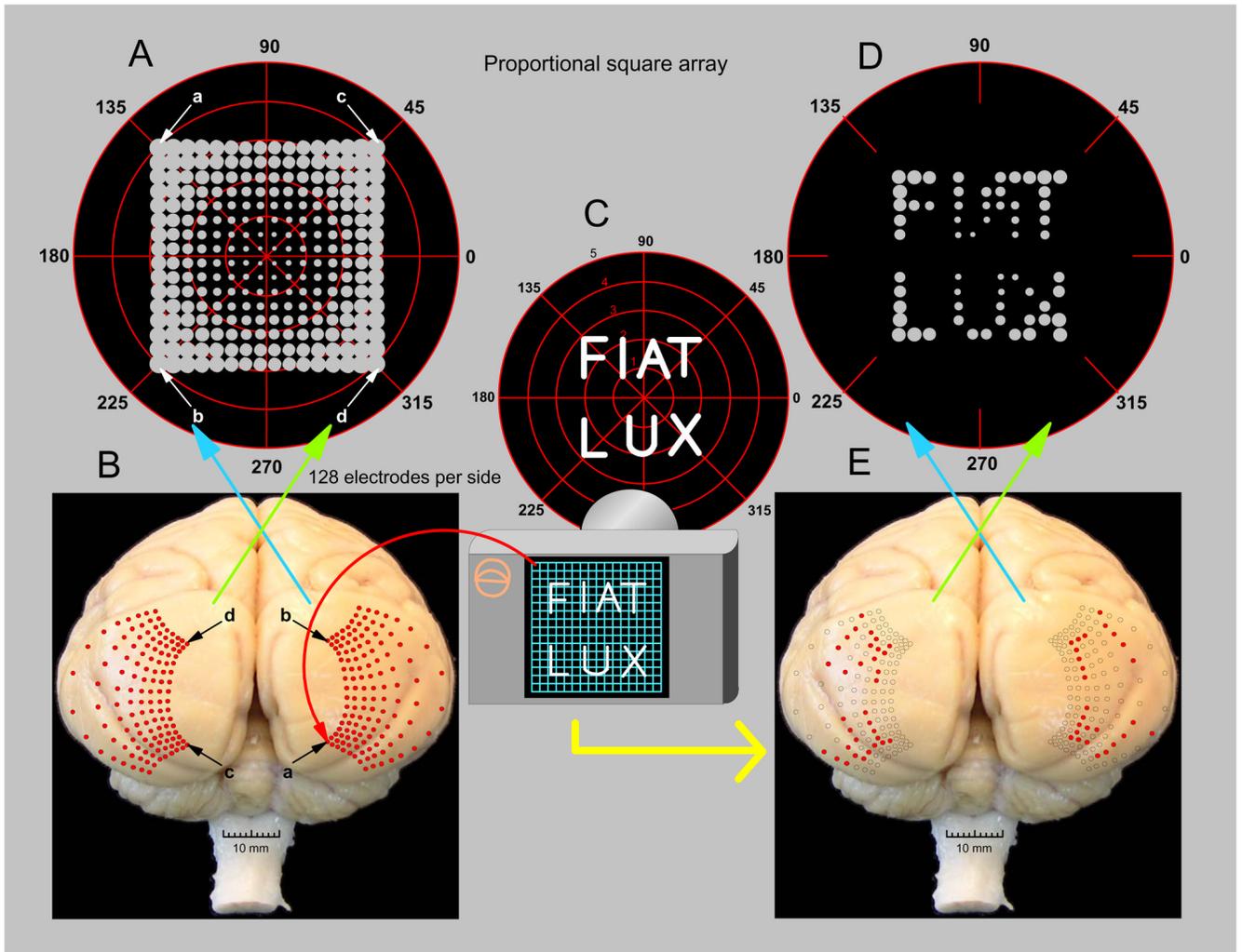


Figure 7.

The presumed effects of electrical stimulation using a proportional square array. In B is shown the rear view of a monkey brain with the electrodes placed bilaterally as had been shown in Figure 6B. When electrical stimulation is applied to all electrodes, as indicated by the red-centered electrodes, it presumably yields the square array of dots shown in A, above. C shows a digital camera that looks at the display FIAT LUX centered in the visual field. The camera is hooked up to a computer and a stimulator arranged to activate the appropriate points in the cortex. The regions activated by the letters are shown by the red-centered dots in E. The resultant image created by the selective activation of the subregions in the camera unit is shown in D.

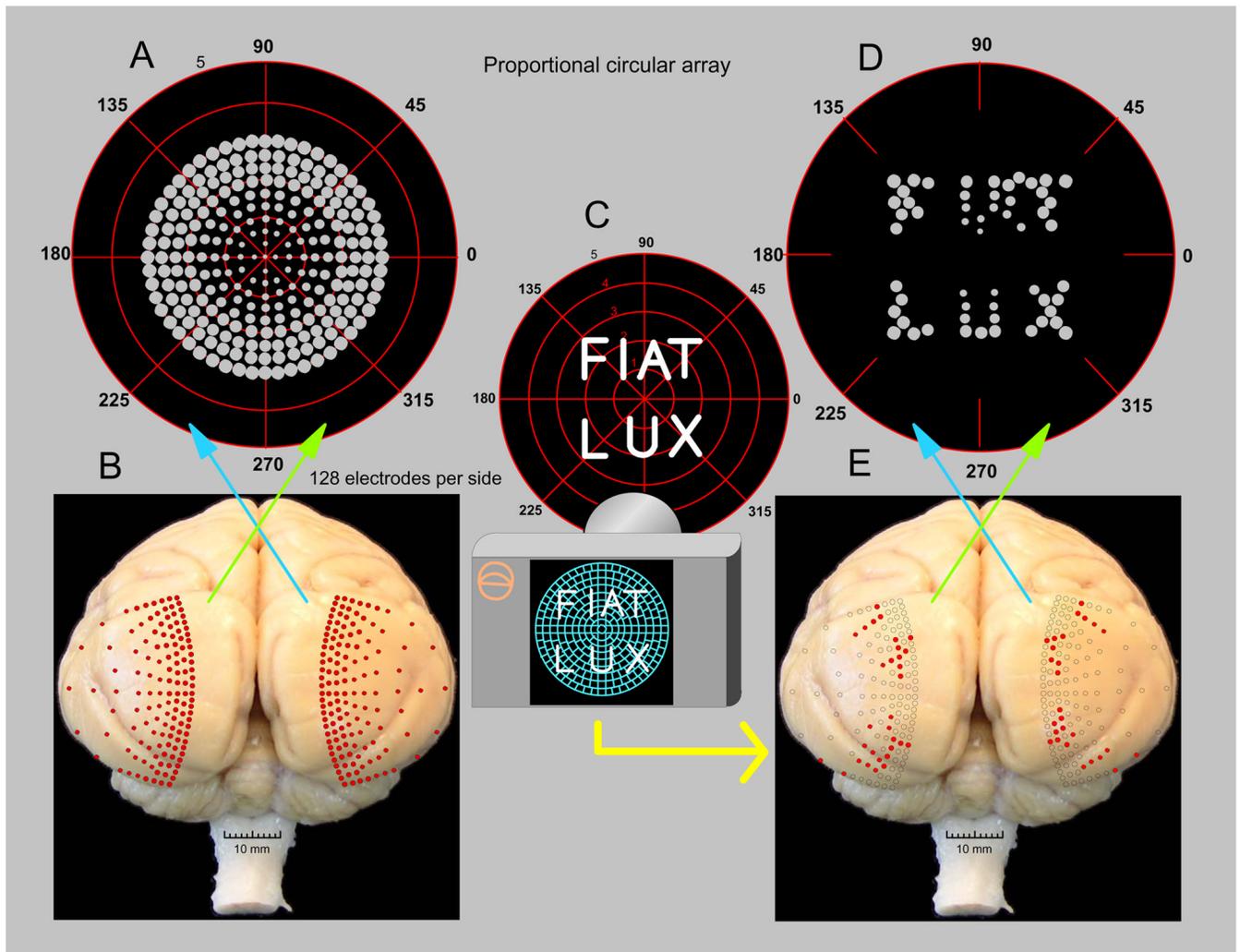


Figure 8. Examination of the percept yielded when a proportional circular array is placed on the cortical surface, shown in B. When all electrodes are activated a radial display comprised of an array of near-perfect circles is produced, shown in A. When the same words, FIAT LUX are presented, shown in C and E, this arrangement yields an image depicted in D.

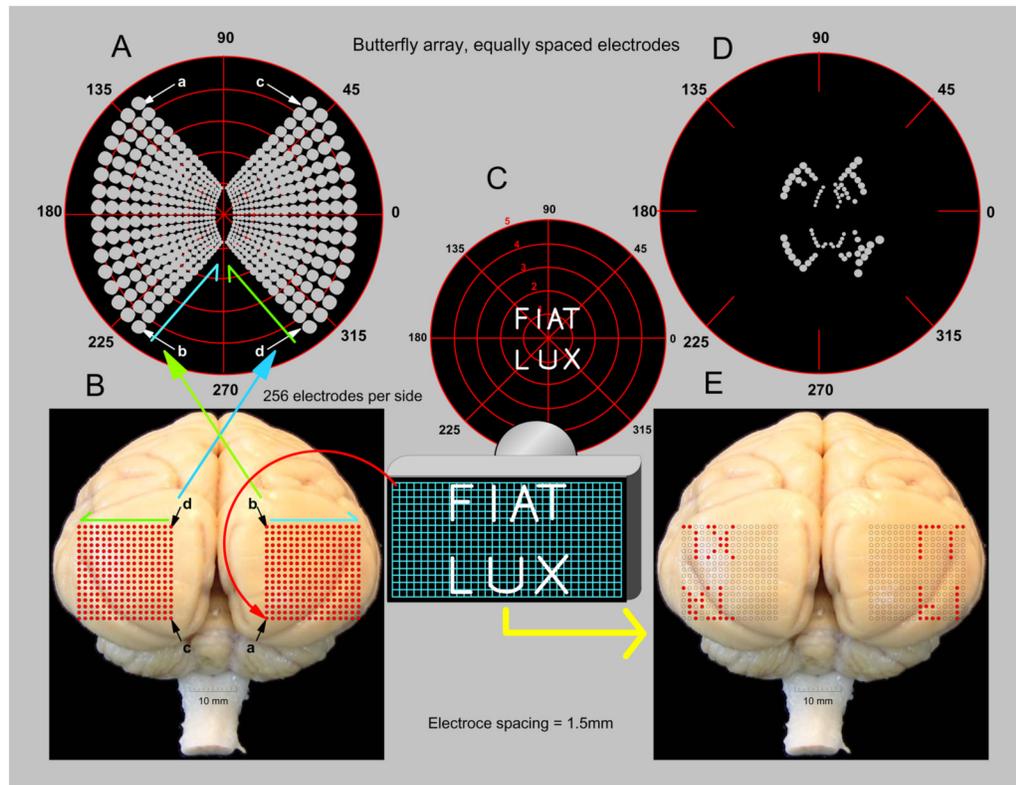
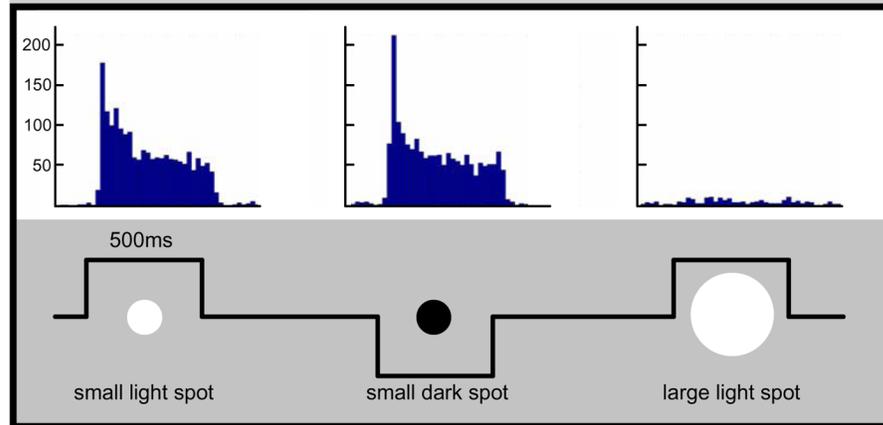


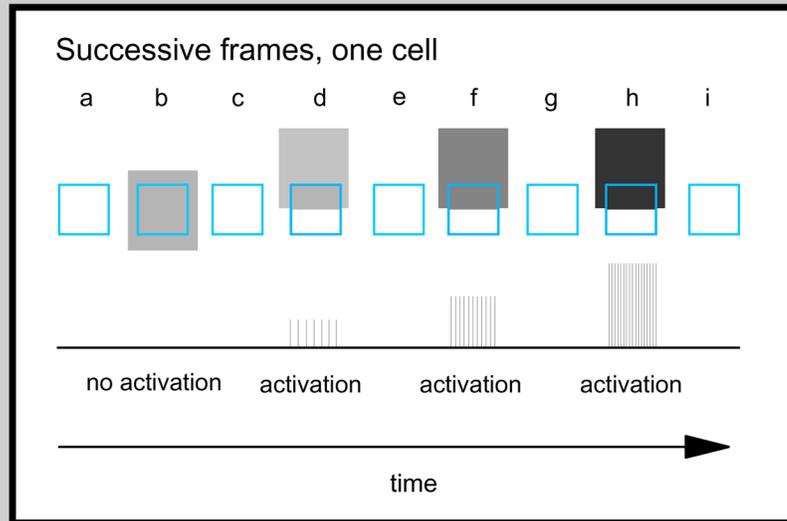
Figure 9.

The images created when equally spaced electrode arrays are placed onto the visual cortex. The arrays consist of 256 elements per hemisphere, each spaced 1.3 mm apart thus making for twice the overall number of electrodes as in the previous figures. When all electrodes are activated, shown in B, the image created is presumed to have the appearance of a butterfly (shown in A) due to the manner in which the visual field is laid out on the cortical surface. Presenting the words FIAT LUX, shown in C and E with the electrodes activated in red, produces a rather distorted image shown in D.

A: Multiunit responses in area V1



B: Intracell comparator



Activation occurs only when a difference in illumination arises within a cell, which makes it equivalent to being an edge detector

Figure 10.

A. Post stimulus time histograms of multiunit data obtained from a fixating alert monkey when the receptive fields of the V1 cells, located in the lower visual field at an eccentricity of 3.2 degrees, were stimulated with a small white spot, a small black spot and a white spot larger than their receptive fields. The data are based on 40 trials for each condition. The small light and dark spots elicited similar responses whereas the large spot elicited no response at all. Standard glass coated platinum/iridium electrodes were used with tip diameters of 1–2 μm and exposed shaft of 10–15 μm , having 40–100 picofarad capacitances. B. The activation of a single cell in the recording system that is set up to have the same number of units as the number of implanted electrodes. Each unit is activated only when a difference in activation occurs *within* its elements, as shown in d. Changes in illumination that affect all elements within a

unit equally, as in b, produce no activation. This arrangement mimicks the basic characteristics of V1 neurons.

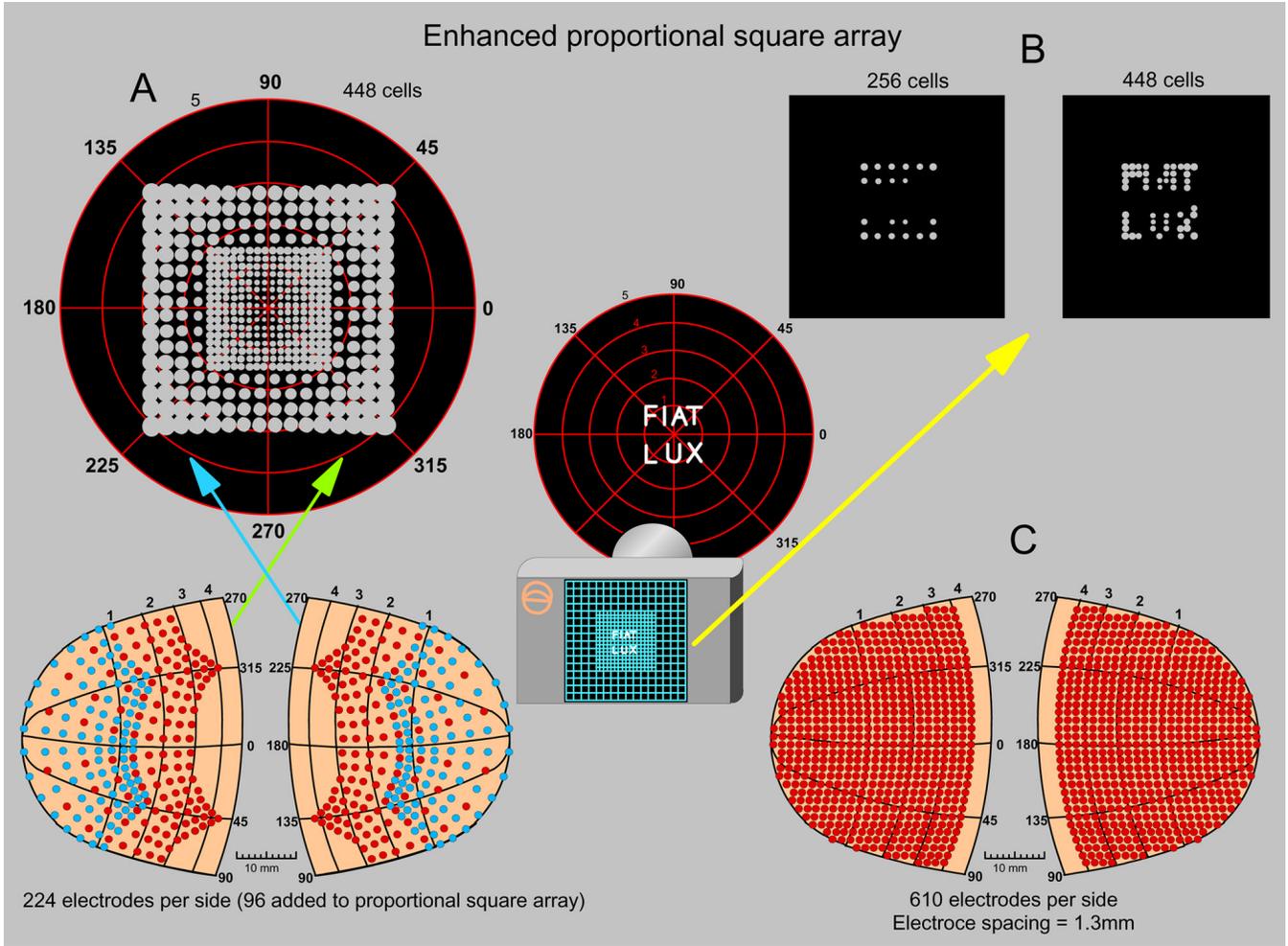


Figure 11.

An enhanced proportional display system. A: Ninety-six electrodes are added to each array (shown in Figures 6 and 7) thereby making for a total of 448 elements. The added electrodes are placed in-between the central 8 by 8 portion of the electrode array shown in Figure 6 yielding a 16 by 16 array. The image elicited by stimulating all sites appears on top. B: The images created by the 256 and 448 element arrays when the words FIAT LUX are confined largely to the central two degrees of the visual field appear demonstrating the higher resolution reaped by the addition of the 96 electrodes. C: Evenly spaced electrode array that allows for proportional activation using a program that corrects for magnification factor. The arrangement of 610 electrodes per side provides approximately the same resolution as the proportional array in A that has 224 electrodes per side.

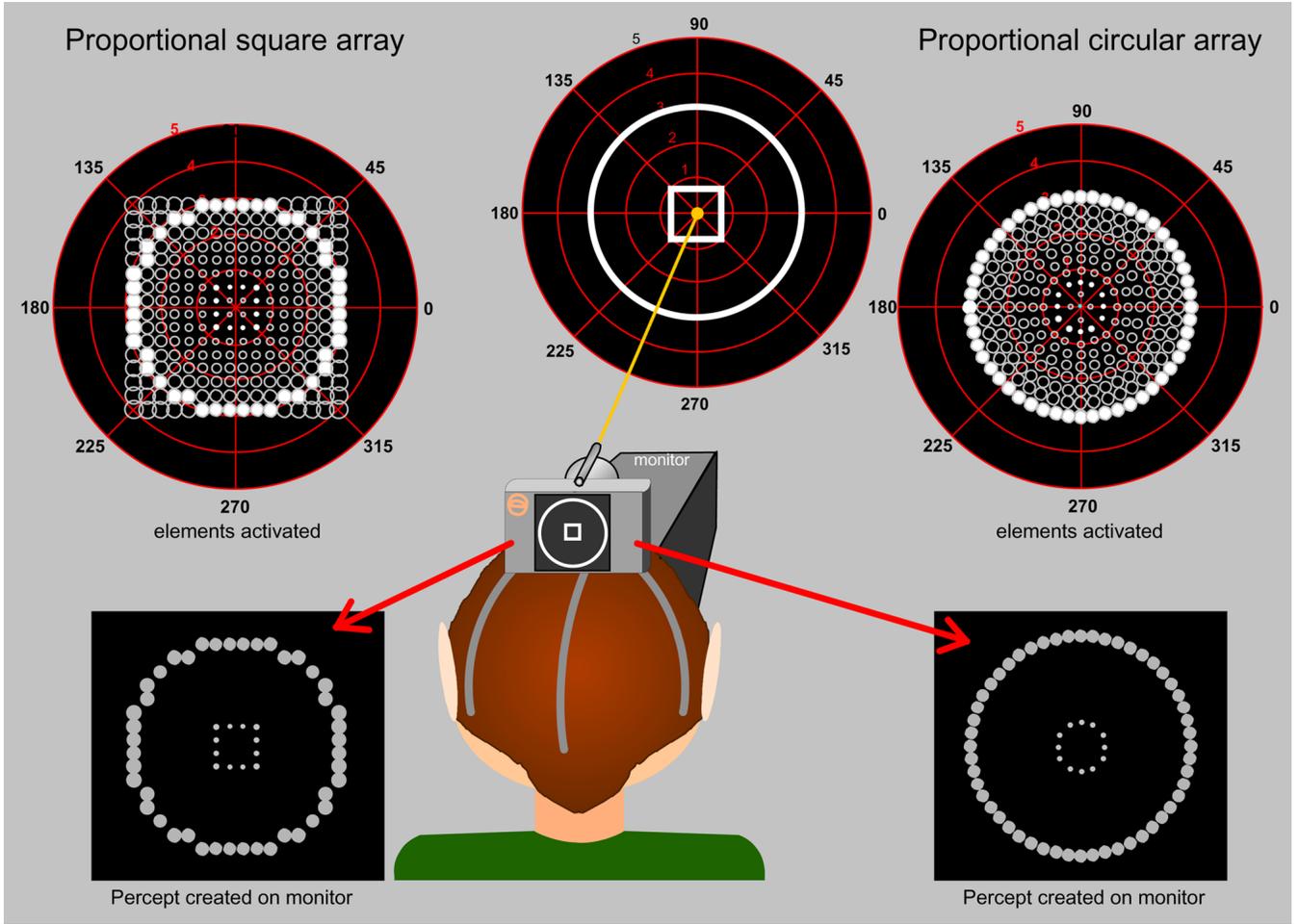


Figure 12. Procedures for mimicking a prosthetic device non-invasively. A video camera is attached to the head with a laser so head movements can be tracked. The image in the camera is converted as in Figure 7 and 8. The resultant image is displayed on a small monitor inside a bezel that is attached to the head, which is the only visual signal provided to the observer. Shown are the layouts for the proportional square and circular arrays when a small central square and a large circle are presented in the visual field. The small square is reproduced well with the proportional array; the large circle is produced extremely well with the circular array.