

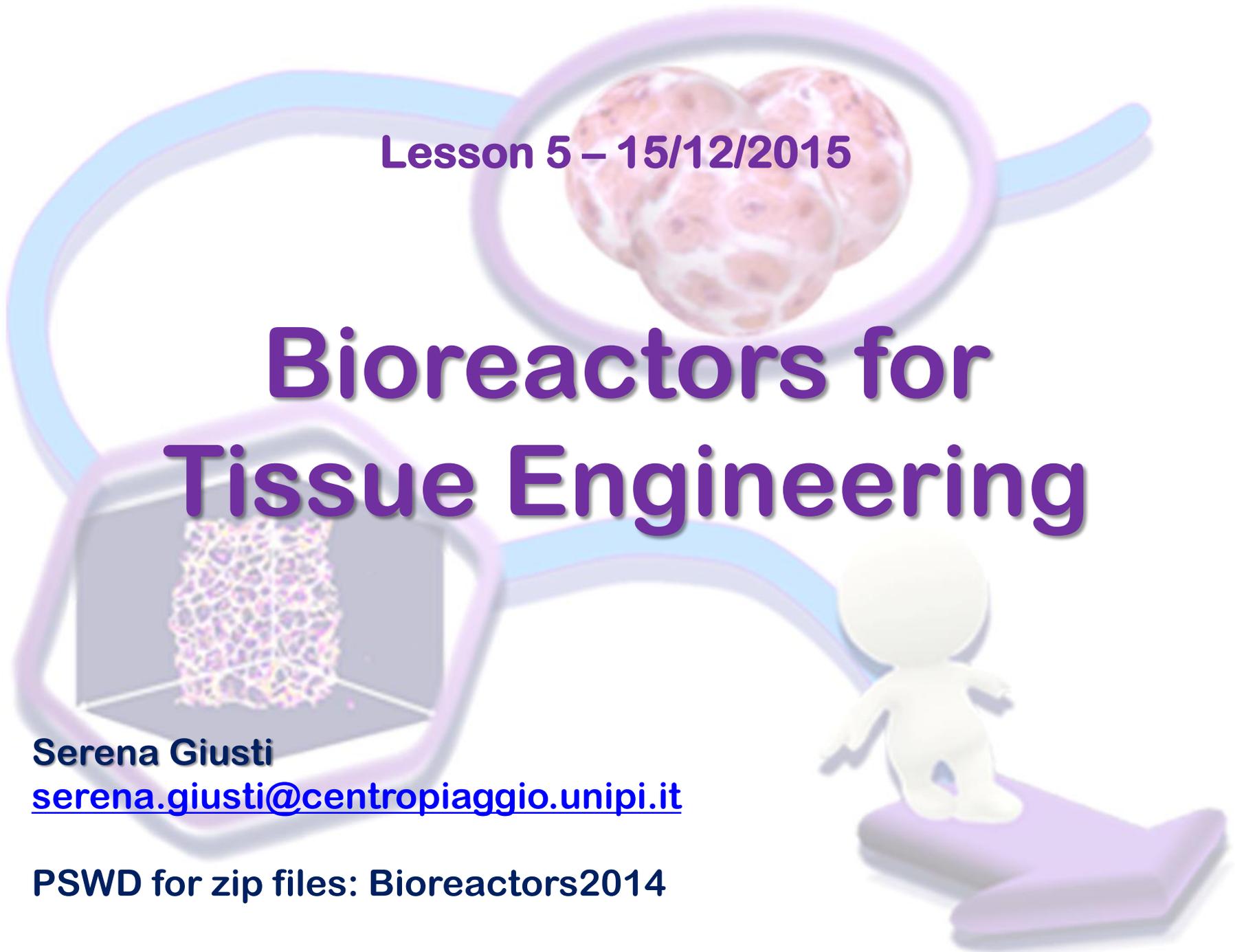
Lesson 5 – 15/12/2015

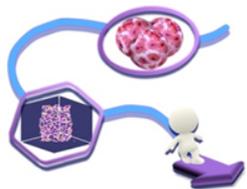
# Bioreactors for Tissue Engineering

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PSWD for zip files: Bioreactors2014





# Connected Cell Culture

## Coltivazione in connessione con altri sistemi/tessuti

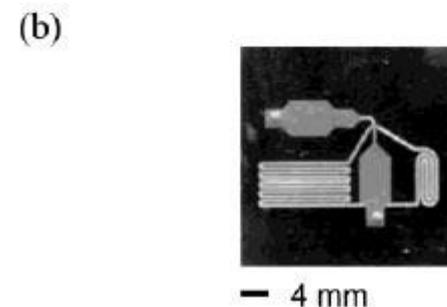
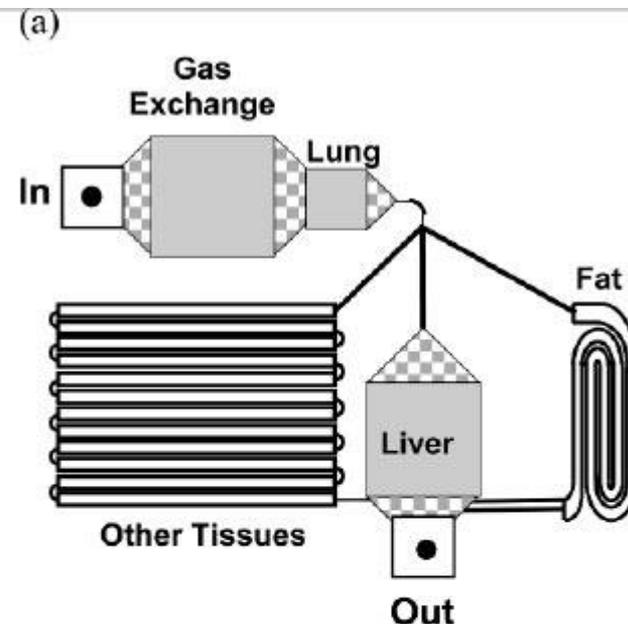
Sono sistemi atti alla valutazione dell'interazione fra i diversi tessuti.

Sono utilizzati in:

1. Studi farmacologici di attivazione o disattivazione di molecole da parte di tessuti (generalmente fegato e tessuti target)
2. Studi di Cross-Talking cellulare
3. Ogni qualvolta si debba investigare l'interazione fra diversi tessuti, senza che questi siano coltivati in diretto contatto l'uno con l'altro (co-cultura)

I bioreattori multi compartimentali o interconnessi sono diversi dalle co-culture!!

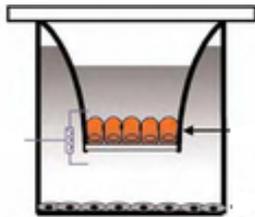
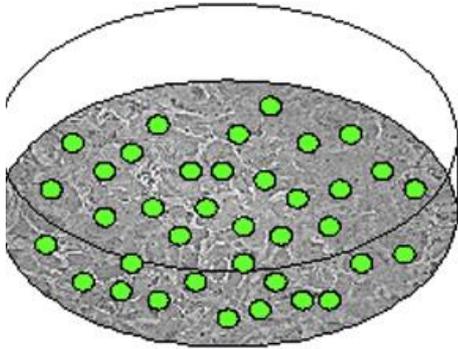
Co-cultura  $\neq$  Riutilizzo del terreno  $\neq$  Coltura connessa



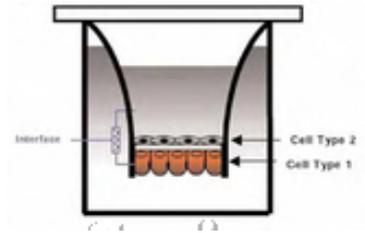
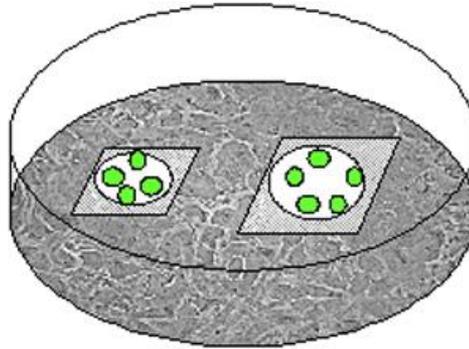


# Connected Cell Culture vs Co-cultures

„contact” coculture



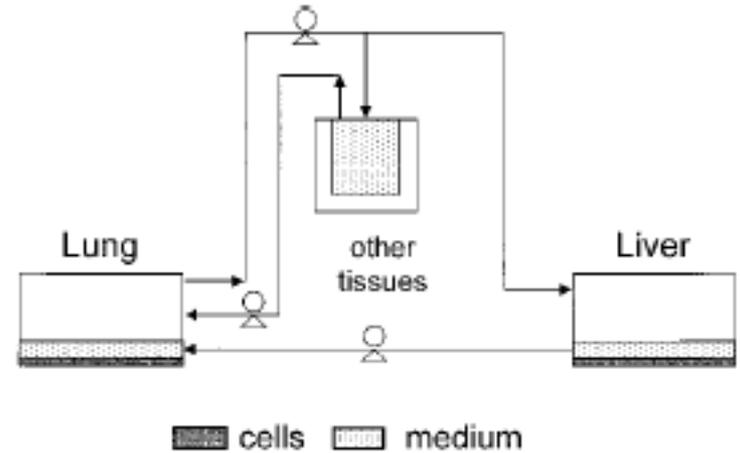
„non-contact” coculture



Co-culture

≠

Conditioned media



Connected culture



# Connected Cell Cultures – PBPK model

- ❑ The **PBPK (Physiologically based pharmacokinetic)** is mathematical modeling technique for predicting the Absorption, Distribution, Metabolism and Excretion (ADME) of synthetic or natural chemical substances in humans and other animal species
- ❑ The PBPK model divides the body into **compartments** representing **organs**, integrating:
  - kinetic
  - thermodynamic
  - anatomical parameters

$$\frac{dQ_i}{dt} = F_i \left( C_{art} - \frac{Q_i}{P_i V_i} \right)$$

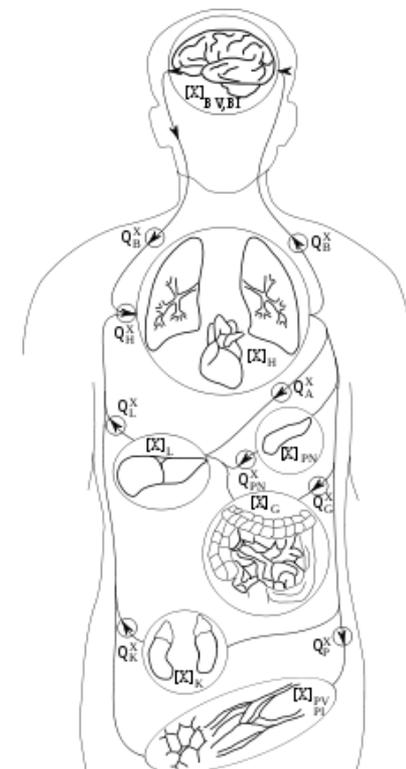
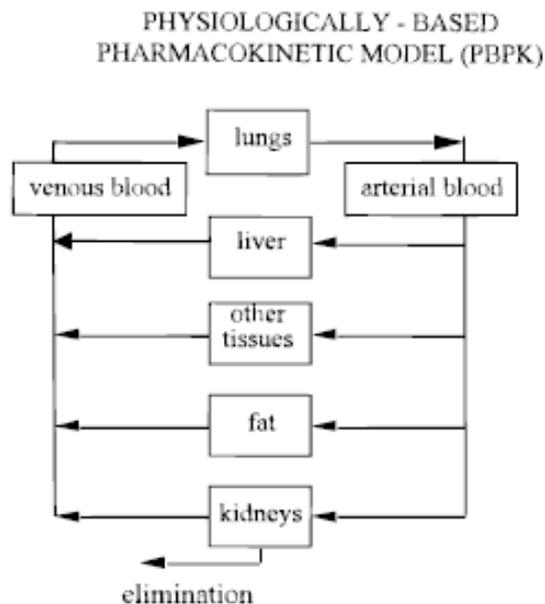
where

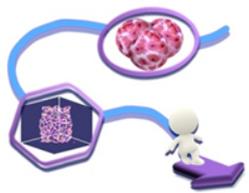
$F_i$  is blood flow,

$C_{art}$  incoming arterial blood concentration,

$P_i$  the tissue over blood partition coefficient

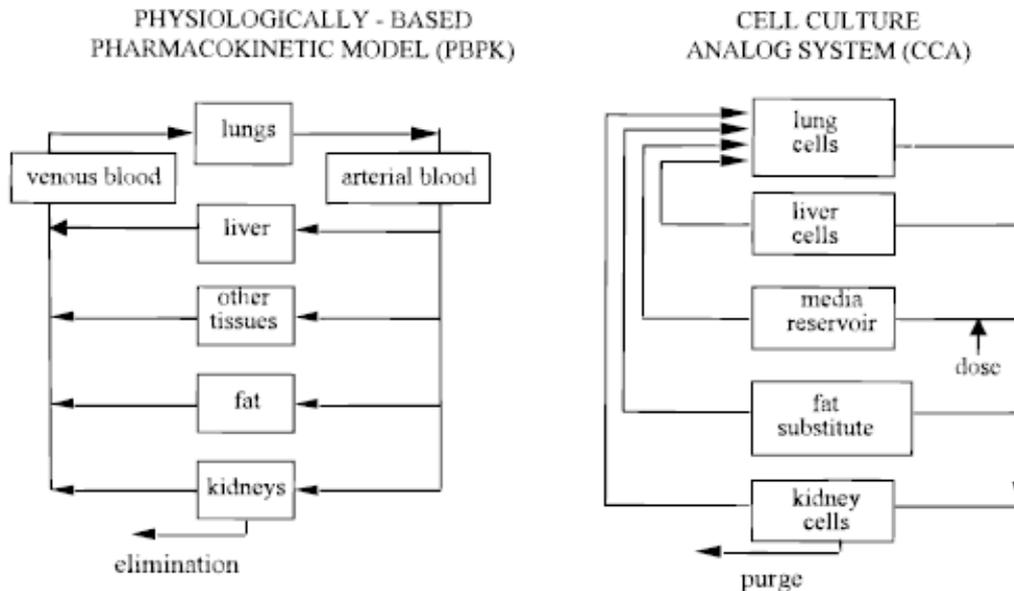
$V_i$  the volume of compartment  $i$ .

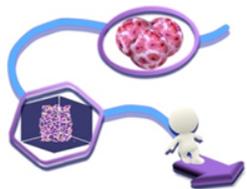




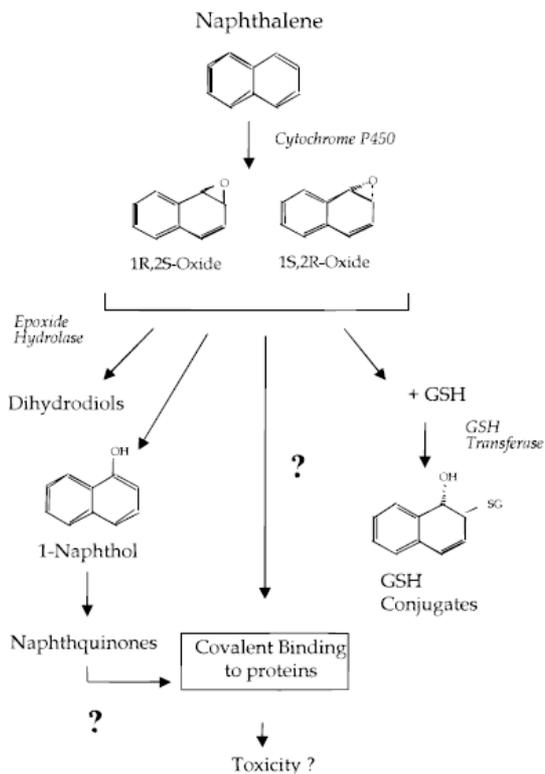
# Connected Cell Cultures – PBPK model

- ❑ The cell culture analogue reactor (CCA) is a physical replica of the PBPK; where the PBPK specifies an organ or tissue compartment, the bioreactor contains compartments with a corresponding cell type.
- ❑ A cell culture analogue of a PBPK can be constructed replacing each PBPK compartment with a biological system that contains the key reactions corresponding to the PBPK kinetic and thermodynamic equations.
- ❑ The device is a **continuous, dynamic system** composed of multiple cell types that interact through a common circulating cell culture medium.





# Case study: naphthalene

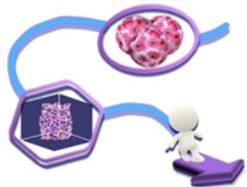


**Naphthalene** has been selected as a model compound for proof-of-concept studies:

*naphthalene metabolism is representative of a broad range of nonnutritive polycyclic aromatic hydrocarbons.*

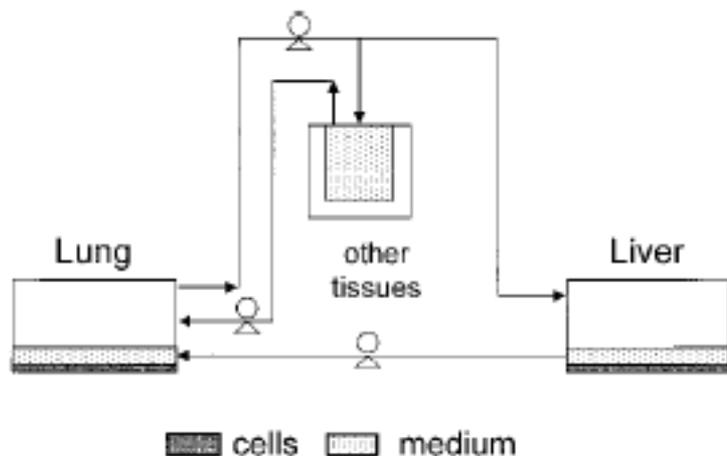
Naphthalene is metabolized in the body by the cytochrome P450 monooxygenase system (CYP450). The primary site of naphthalene metabolism is the liver, where it is metabolized into naphthalene oxide enantiomers.

The primary target of naphthalene is the non-ciliated bronchial endothelial cells.



# Key points

- ❑ In the CCA is used only a single cell type in each “organ”: L2 cells in the lung compartment and H4IIE cells in the liver compartment
- ❑ the “organ” compartments contain the key enzymes of the PBPK model.
- ❑ The CCA has advantages over other in vitro systems, thus to allow quantitative measurements



- **A 300 mg/kg** dose of naphthalene (dissolved in methanol)

Results:

- Depletion of intracellular glutathione in the L2 cells 6 h after dosing.
- **Depletion in liver cell glutathione** is shown after 4h from dosing, but not at 6h
- *Controls for the experiments* were flasks of the same cell types prepared in the same way as cells in the reactor but kept in an incubator during the reactor run, and different CCA configurations.



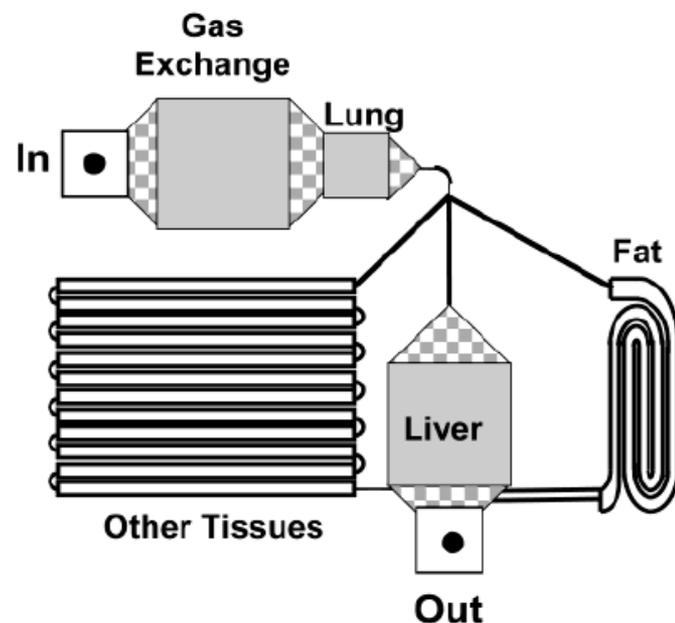
# The «micro» CCA-PBPK

- A microscale CCA ( $\mu$ CCA), also called “**animal-on-a-chip**”, may be constructed to better mimic the appropriate physiological scale (e.g., characteristic dimensions of tens of microns).

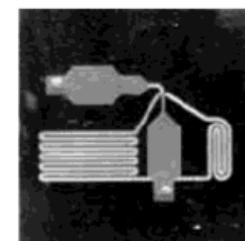
compartment	calculated residence time (s)	measured residence time (s)
lung	2	2
liver	28	38 ± 10
fat	134	120 ± 12
other-tissue	207	200 ± 20

The culture medium first entering the system through the inlet passes through to the “lung”, after which;

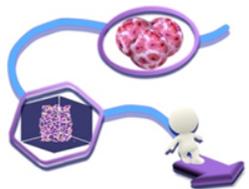
- 9% to the fat chamber
- 25% to the liver chamber
- 66% “other tissue” chambers



(b)



— 4 mm



# The «micro» CCA-PBPK

Cell cultures:

The lung and liver chambers were then coated with Matrigel at  $100 \mu\text{g}/\text{cm}^2$  density:

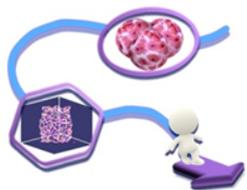
- $40 \mu\text{L}$  of L2 at  $2 \times 10^5$  cells/mL were plated into lung chamber (8000 cells)
- $40 \mu\text{L}$  of HepG2/C3A or H4IIE at  $1 \times 10^6$  were seeded into liver chamber ( $40000$  cells) **5 hep : 1 Lung**

Flow at  $2 \mu\text{L}/\text{min}$ .

NB. In the «fat» and «other tissues» compartments there were NO CELLS

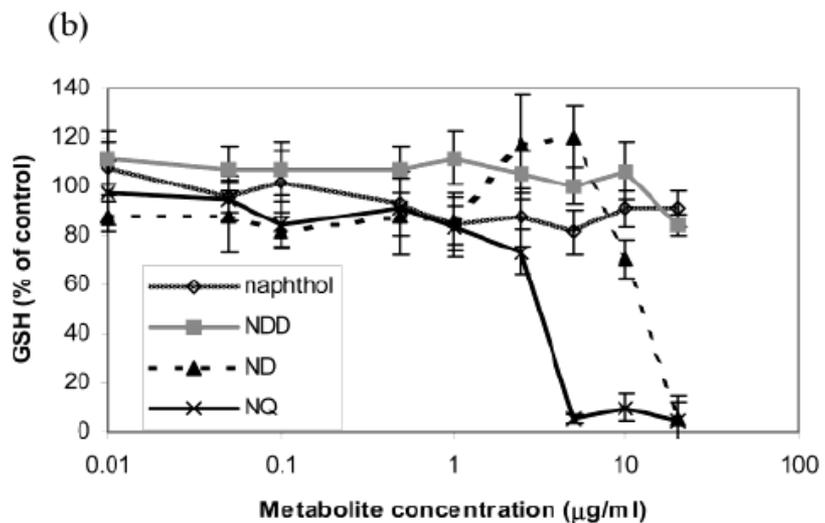
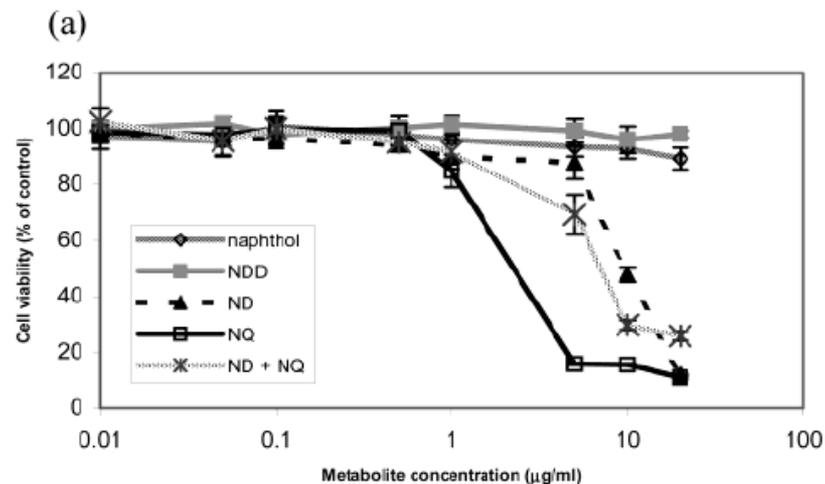
The “fat” compartment is added to a previous three chamber iCCA system to better mimic the fluid distribution in rapidly and slowly perfused organs and more accurately predict the response of animals to chemicals.

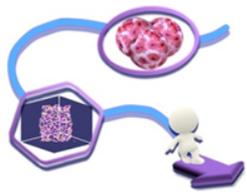




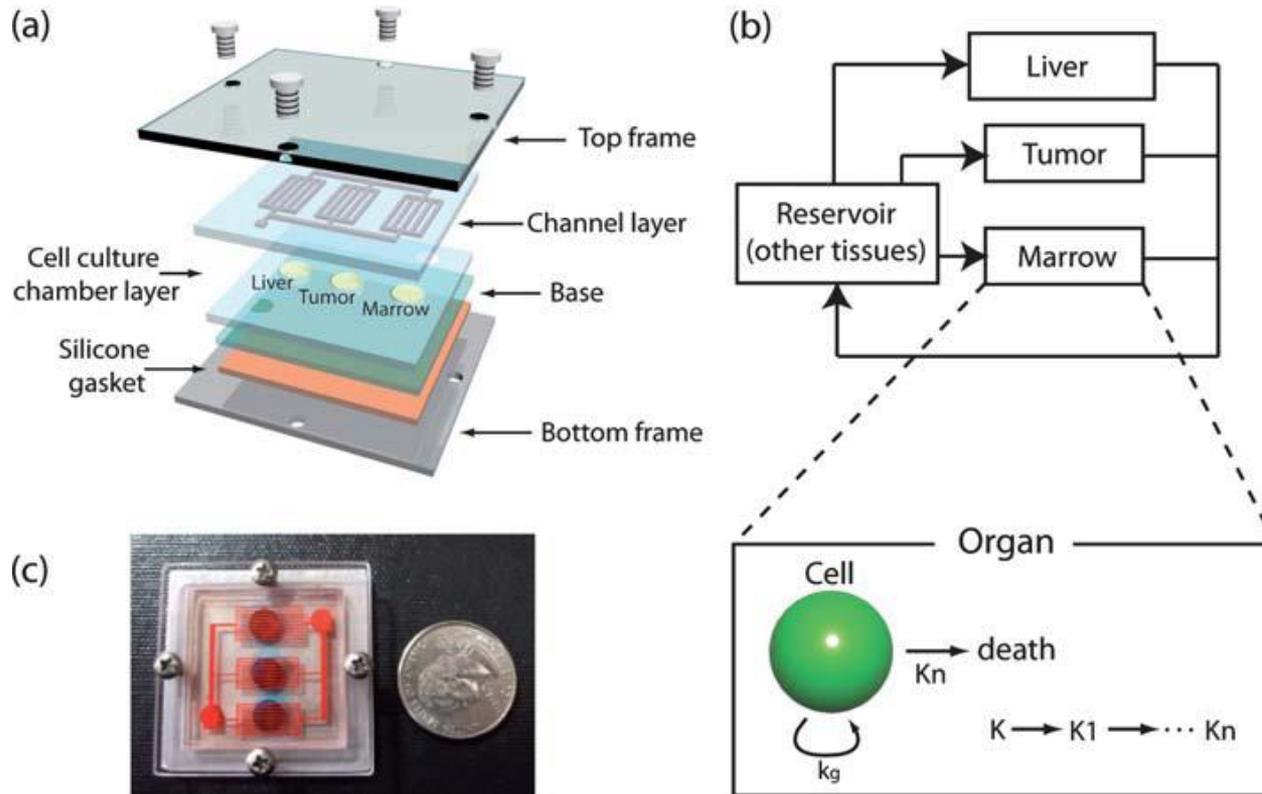
# Results $\mu$ CCA

- ❑ naphthalene diol and naphthoquinone significantly decreased cell viability
- ❑ Calcein stain revealed that naphthoquinone drastically changed the morphology of L2 cells
- ❑ Unlike L2 cells, the morphology of C3A did not change upon being exposed to naphthoquinone
- ❑ GSH in both cell lines, L2 and C3A, decreased with increasing concentration of naphthoquinone
- ❑ After 6 h experiment, there was no intracellular GSH depletion in L2 observed in the L2-blank and L2-L2 combinations





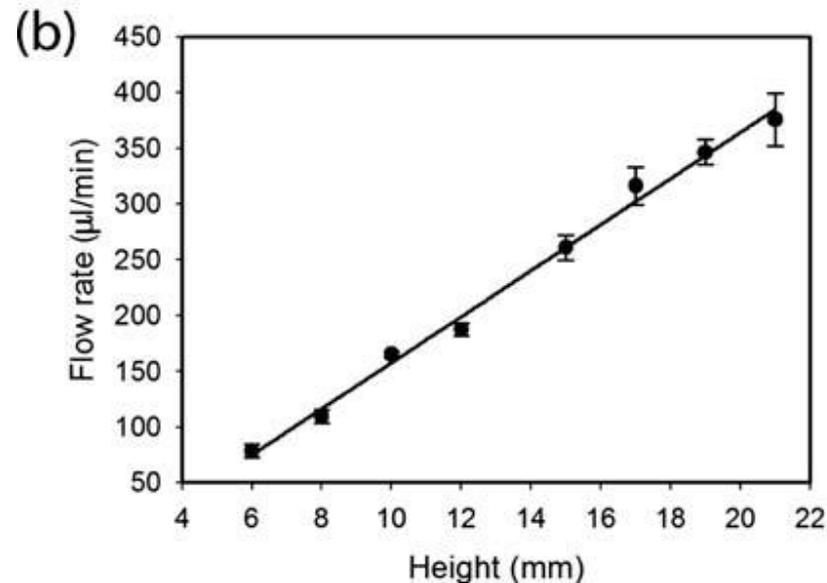
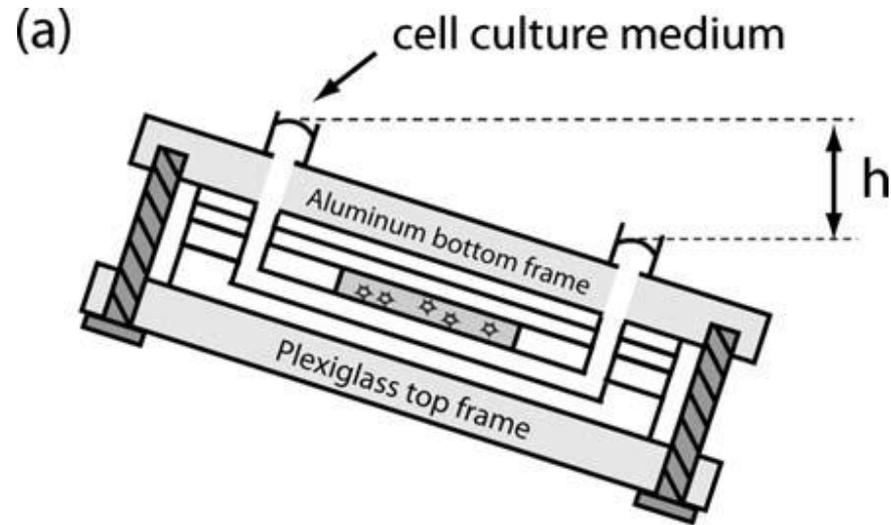
# A new application for $\mu$ CCA



(a) A schematic of device components. A fluidic channel layer and a cell culture chamber layer are superimposed and sealed by top and bottom frames. A silicone gasket and a polycarbonate base are inserted for sealing. (b) A corresponding PBPK model, with the liver, tumor and marrow compartment. Below is a PD model for cell death in each compartment. Although not drawn explicitly, a PD model for each compartment exists separately, and the 'organ' can be the liver, tumor or marrow. (c) A picture of the assembled device. A red dye was used for visualization of channels, and a blue dye was mixed with alginate.



# A new application for $\mu$ CCA



Although microfluidic systems have a great potential in enhancing the drug development process, actual applications of microfluidic systems in medical or life science area have been limited.

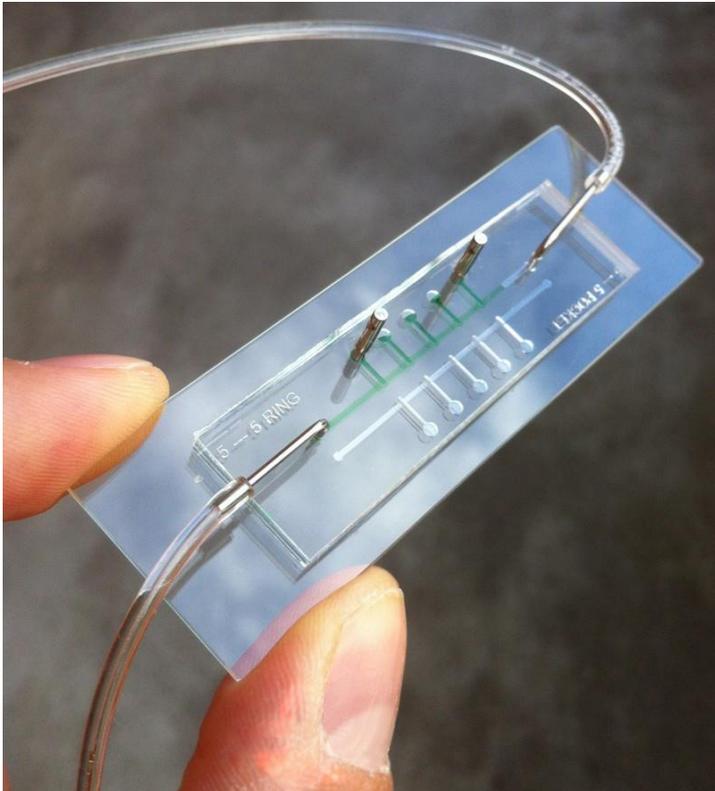
- current microfluidic devices require specialized skills for fabrication and operation, which makes it difficult to be used by non-experts.
- microfluidic cell cultures need more indepth study, such as maintenance of sterility, formation of air bubbles, the effect of shear stress on cells, and the «edge-effect»

The use of gravity-induced flow eliminates the need for a pump, and prevents formation of air bubbles.

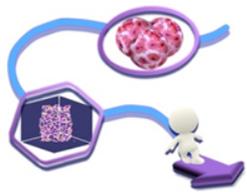


# Milli VS Micro scaled systems

**Micro-fabricated micro-fluidic cell culture systems** are highly cited, but remain a **niche research tool** with **several intrinsic limitations**



- **Cell culture surface is ~ 0.5-0.8 mm<sup>2</sup> and seeded with few thousand cells**
- **The surface/volume ratio is extremely high** giving rise to
  - **High wall shear stresses** having **detrimental effects on many cell types** (e.g. hepatocytes)
  - **Edge-effect** with **peripheral cells** exhibiting **increased cytoskeletal tension** and **altered viability/functionality** than central ones
  - **Air bubbles** causing **fluid pattern alterations**
- **PDMS to enhance oxygen transport**
  - **Adsorb small hydrophobic molecules** (nutrient/ligand depletion, exp. artefacts)
- **Very difficult to handle, assemble and use**



# Milli VS Micro scaled systems

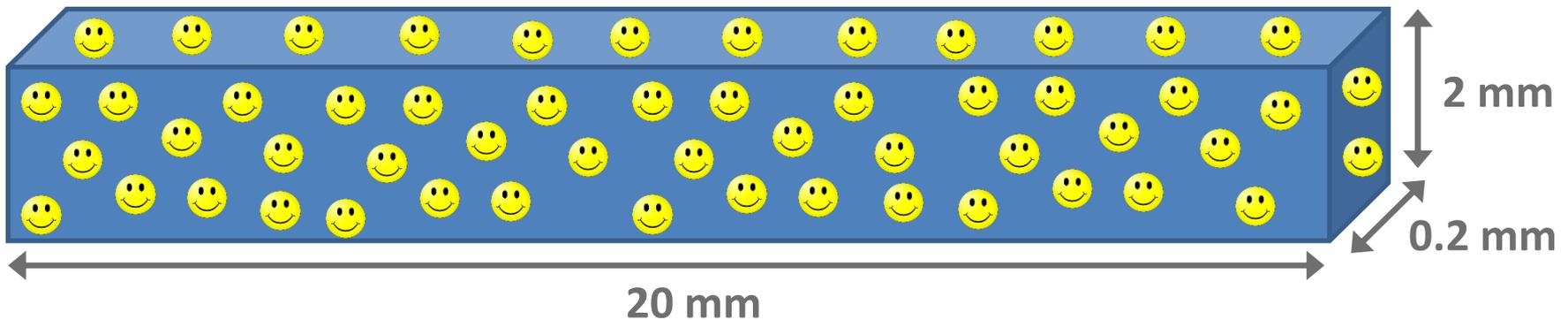
**Micro-fluidic systems also require the translation of conventional experimental methods and protocols established at the milli-scale using standard cell culture supports (e.g. 24 MW plate).**

Micro-Fluidics	Milli-Fluidics
Low shear by reduction of flow rate	<b>High flow rates with low shear</b>
Low nutrient turnover	<b>High nutrient turnover</b>
High surface to volume ratio	<b>Low surface to volume ratio</b>
Fiddly to assemble	<b>Easy to assemble</b>
Presence of air bubbles	<b>No air bubbles</b>
<b>Low fluid volumes, saving on reagents</b>	Higher volumes of media and reagents
<b>Easy quantification of cell products</b>	Cell products may be harder to quantify



# Milli VS Micro scaled systems

Surface of hepatocyte-laden construct in equilibrium with maximum  $c_{O_2}$  (i.e. 0.21 mM)

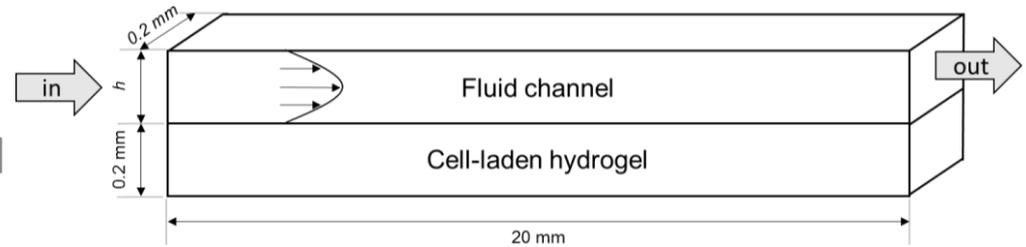


**Not implementable in real fluidic systems as it requires very high medium flow rates that damage cells due to high shear stresses**

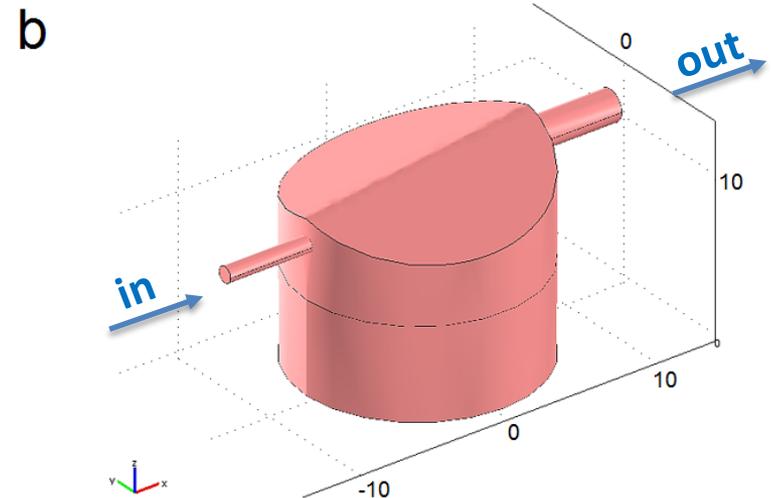
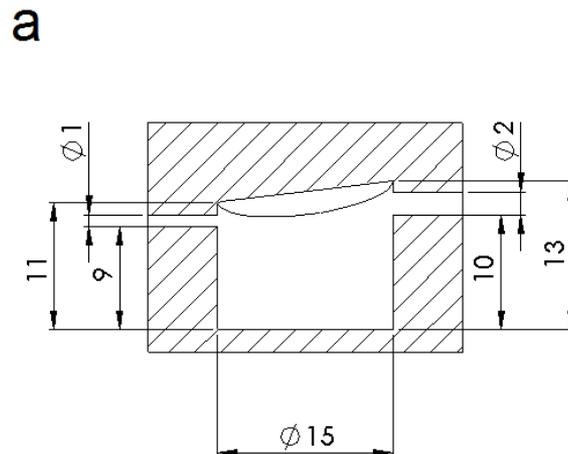


# Milli VS Micro scaled systems

Simple parallelepiped fluidic channel



- ✓ PDMS milli-molded culture chamber
- ✓ Shape and dimensions similar to those of a 24 MW plate well
- ✓ Suited for implementing multi-organ in-vitro models to study organ cross-talk through inter-connected cell cultures



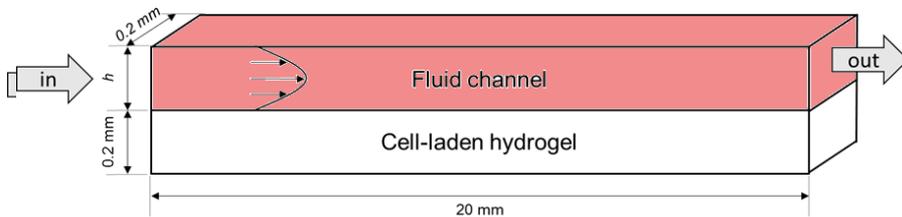


# Milli VS Micro scaled systems

Steady-state multi-physics model which couples oxygen mass transport and consumption to fluid dynamics (solved in COMSOL Multiphysics 3.5a)

## Fluid domain

(O<sub>2</sub> convective transport, no consumption)

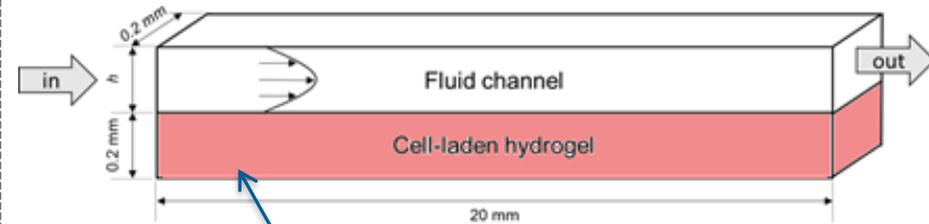


$$v_{in} = \frac{\tau_w \cdot h}{6\eta}, \quad \tau_w = 15 \mu\text{Pa}$$

- $h = 0.2 \text{ mm} \rightarrow v_{in} = 7.1 \times 10^{-7} \text{ m/s}$
- $h = 2 \text{ mm} \rightarrow v_{in} = 1.6 \times 10^{-6} \text{ m/s}$

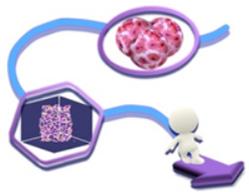
## Hydrogel construct domain

(O<sub>2</sub> diffusion and consumption)



$$R = \frac{V_{max} \cdot c}{K_m + c} \cdot \delta(c > c_{cr})$$

- **0.2 mm** thick gel
- **$7 \cdot 10^7$**  hepatocytes/mL

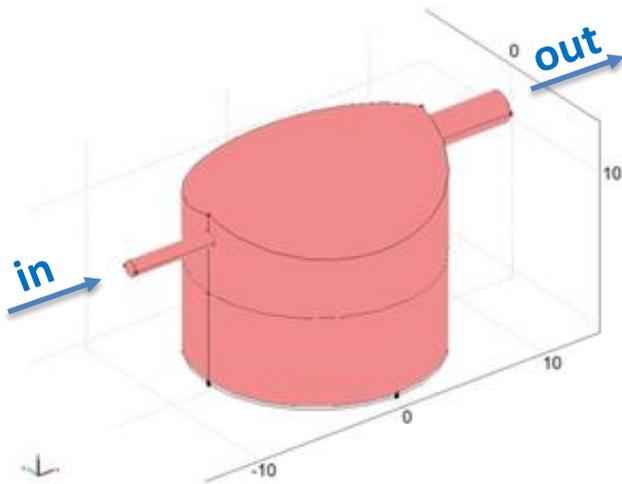


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## Fluid domain

(O<sub>2</sub> convective transport, no consumption)



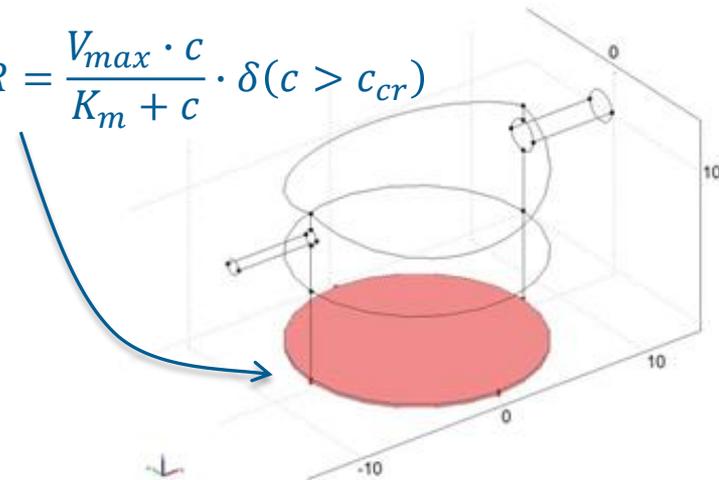
$$v_{in} = 3.82 \times 10^{-3} \text{ m/s} = 180 \text{ } \mu\text{L/min}$$

(from experimental results)

## Hydrogel construct domain

(O<sub>2</sub> diffusion and consumption)

$$R = \frac{V_{max} \cdot c}{K_m + c} \cdot \delta(c > c_{cr})$$

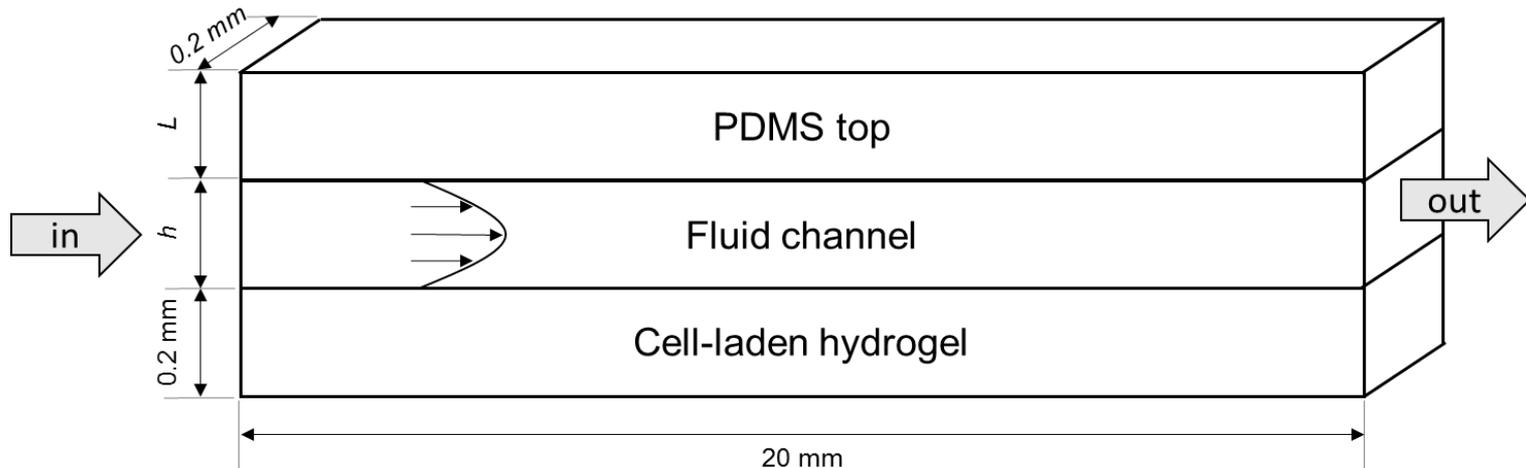


- **0.2 mm** thick gel
- **7·10<sup>7</sup>** hepatocytes/mL



# Milli VS Micro scaled systems

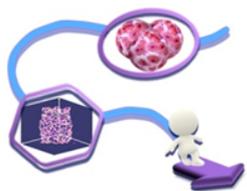
Most **micro-fluidic cell culture devices** are made of **PDMS**, a **gas permeable elastomer**, in order to **enhance oxygen mass transport**



$$N_{O_2,PDMS} = K_{O_2} \left( p_{O_2} - \frac{c_{O_2}}{K_{H,O_2}} \right)$$

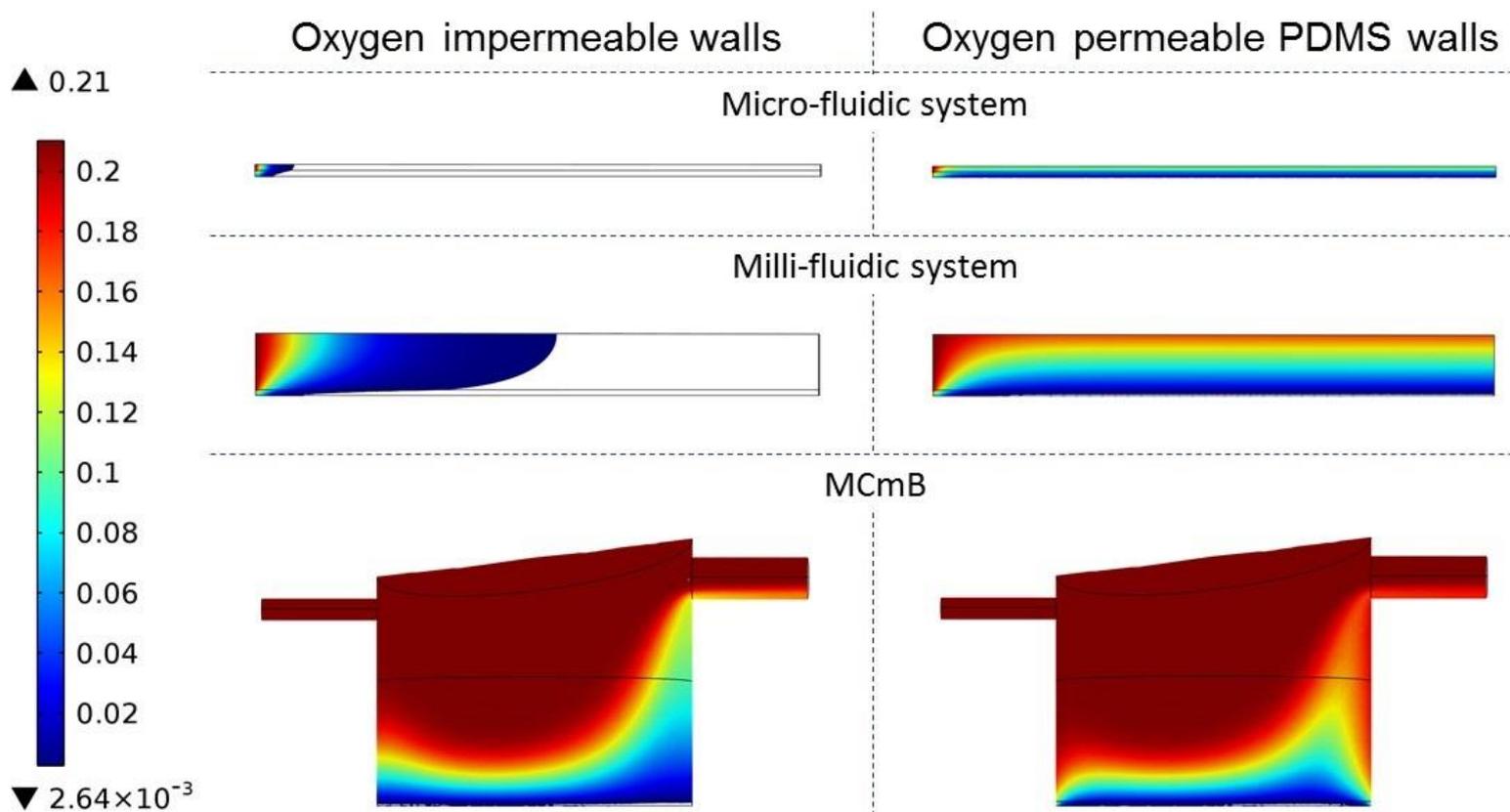
$$K_{O_2} = P_m/L$$

- **Micro-fluidic systems** --> **L = 5 mm** (typical)
- **MCmB** --> **L = 6 mm**



# Milli VS Micro scaled systems

## Oxygen concentration (mM)





# Milli VS Micro scaled systems

Modelled System	Micro-Fluidic		Milli-Fluidic		MCmB	
	OIW	OPW	OIW	OPW	OIW	OPW
Viable cells (%)	2.6	100	17.8	78	60.5	85.4

**OIW:** oxygen impermeable walls; **OPW:** oxygen permeable walls

**O<sub>2</sub> diffusion through PDMS enhances the viability of encapsulated hepatocytes, especially in micro-fluidic systems (PDMS over-reliance\*)**

\* **Micro-fluidic cell culture devices** rely entirely on oxygen diffusion through the walls of the device, and cannot be manufactured from oxygen-impermeable materials, such as tissue culture plastic or glass



# Milli VS Micro scaled systems

Configuration	Flow Rate (m <sup>3</sup> /s)	R <sub>in</sub> (mol/s)	Cell Number	R <sub>cons</sub> (mol/s)	R <sub>in</sub> /R <sub>cons</sub>
Micro-fluidic	$2.84 \times 10^{-14}$	$1.42 \times 10^{-13}$	$5.60 \times 10^4$	$4.48 \times 10^{-13}$	0.3
Milli-fluidic	$6.40 \times 10^{-13}$	$3.20 \times 10^{-12}$	$5.60 \times 10^4$	$4.48 \times 10^{-13}$	7.1
MCmB	$3.00 \times 10^{-9}$	$1.50 \times 10^{-8}$	$2.47 \times 10^6$	$1.98 \times 10^{-11}$	758.3

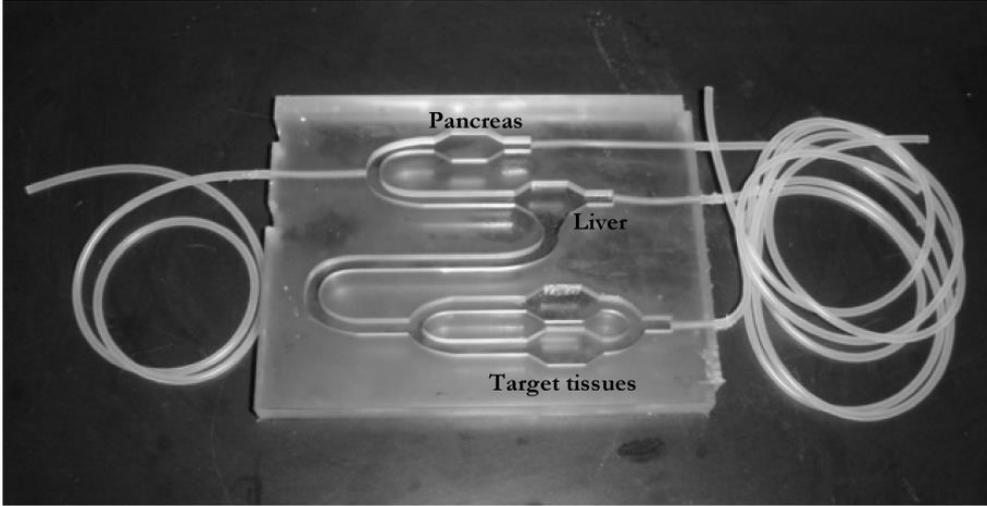
$R_{\text{cons}}$  = glucose cons. rate  $\times$  cell number;  $R_{\text{in}}$  = glucose conc. (5 mM)  $\times$  medium flow rate

**Micro-fluidic systems may suffer from depletion of nutrients other than oxygen if cell densities are increased to better approximate physiological values, while milli-fluidic systems have sufficient mass flow rate to ensure an acceptable  $R_{\text{in}}/R_{\text{cons}}$  ratio**

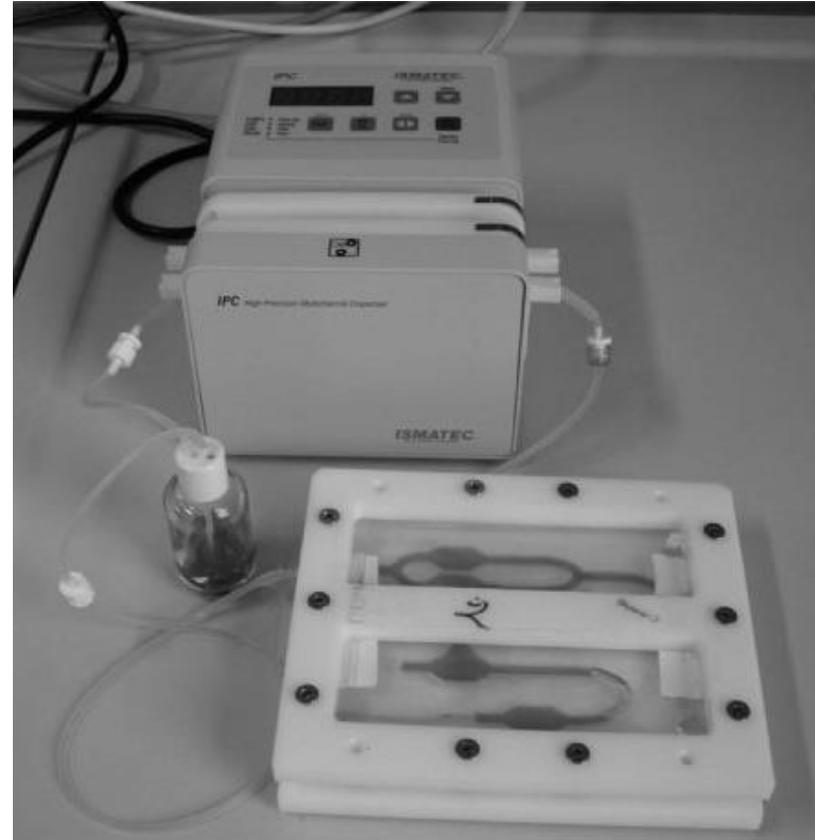
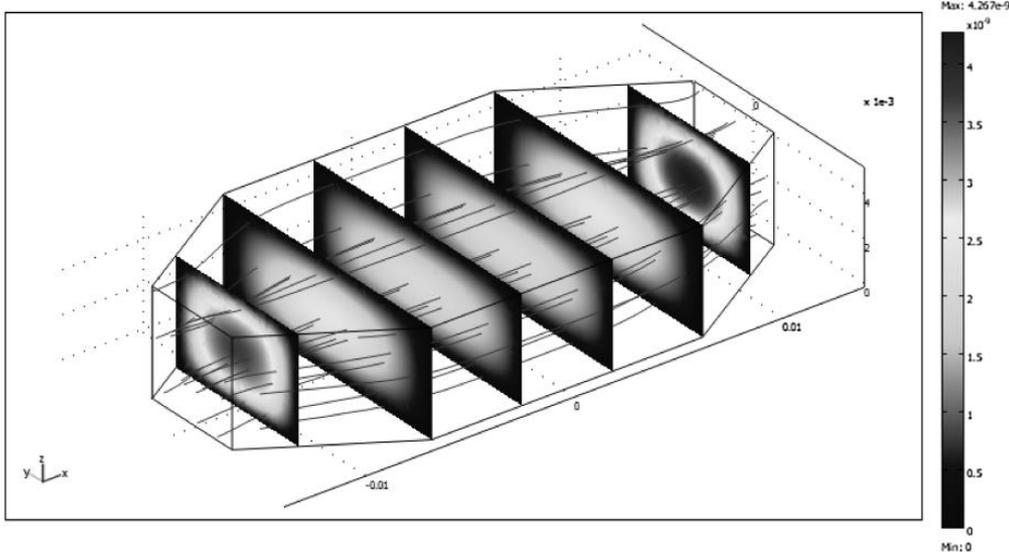


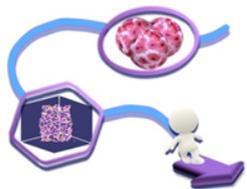
# The MCB

a



b

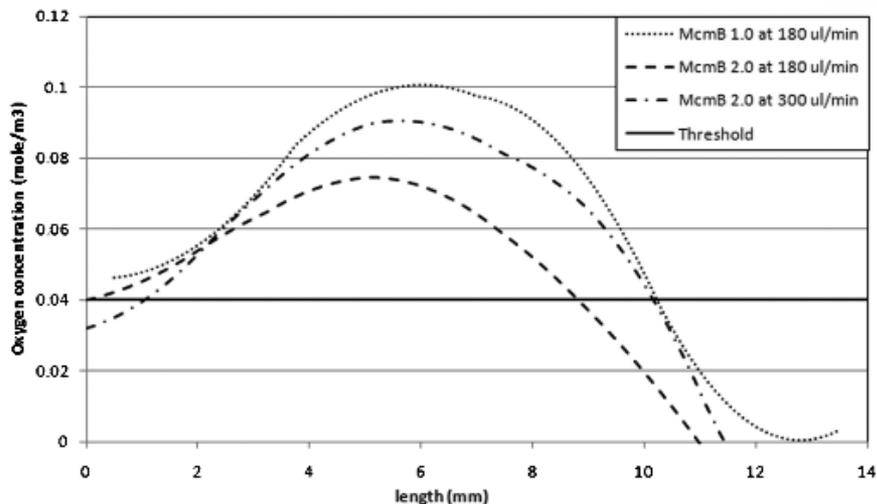
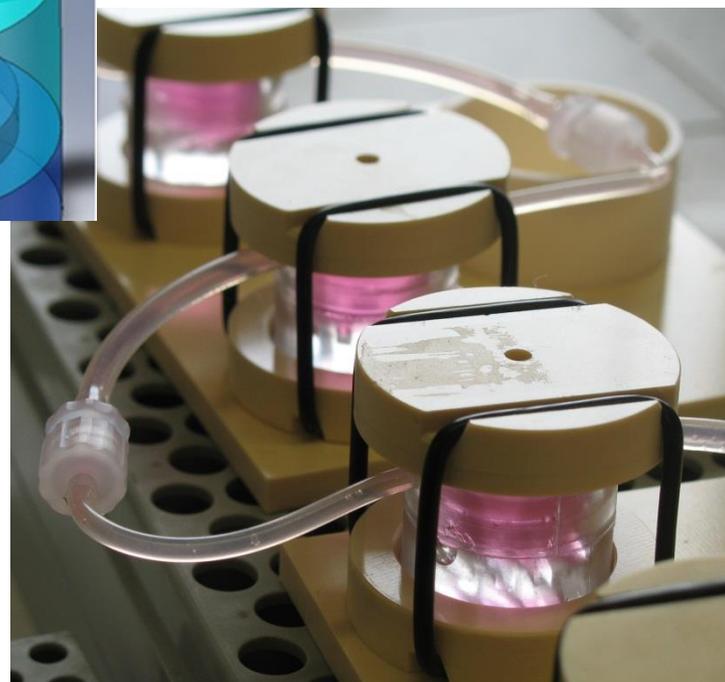
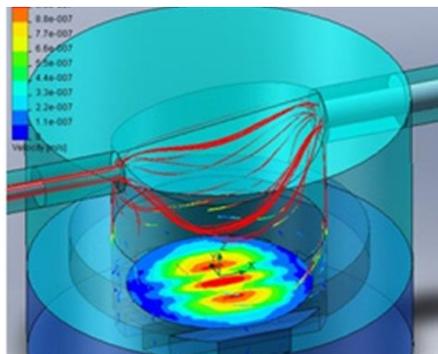


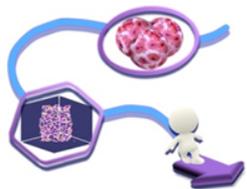


# The MCmB

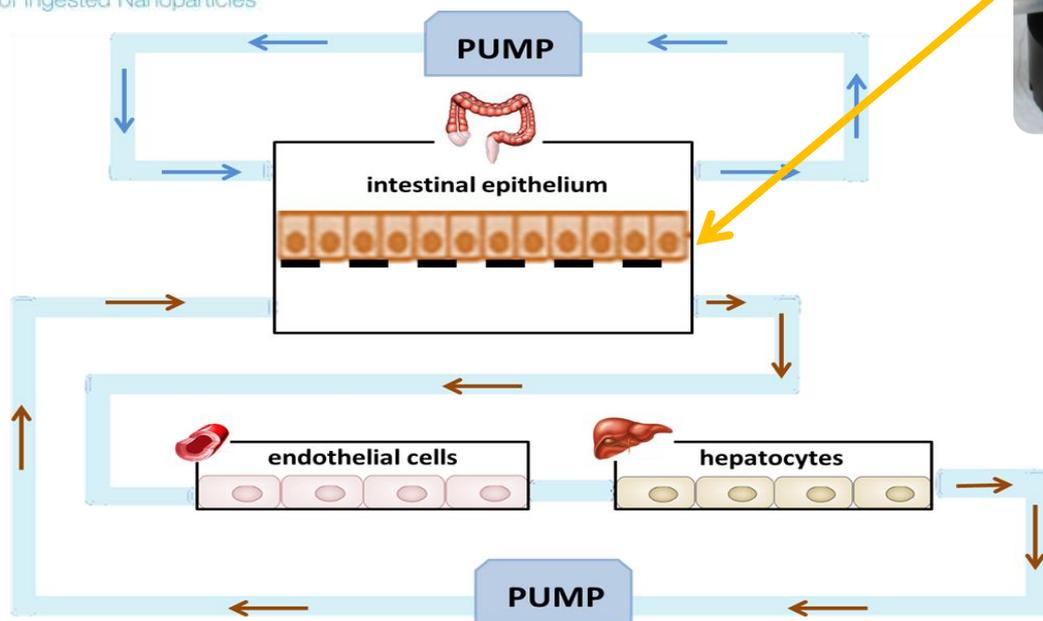
The QuasiVivo<sup>®</sup> (Kirkstall Ltd, UK) is a modular chamber which can be linked to other chambers for high throughput multi-compartment experiments

- High flow rates
- Low shear stress
- Connected cultures

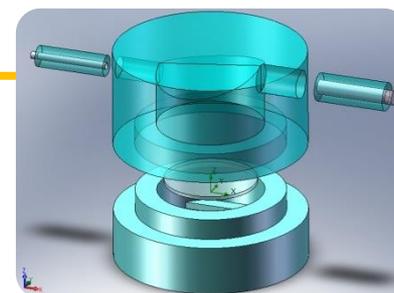




# Connected Cell Culture



2 flow membrane bioreactor with realtime TEER



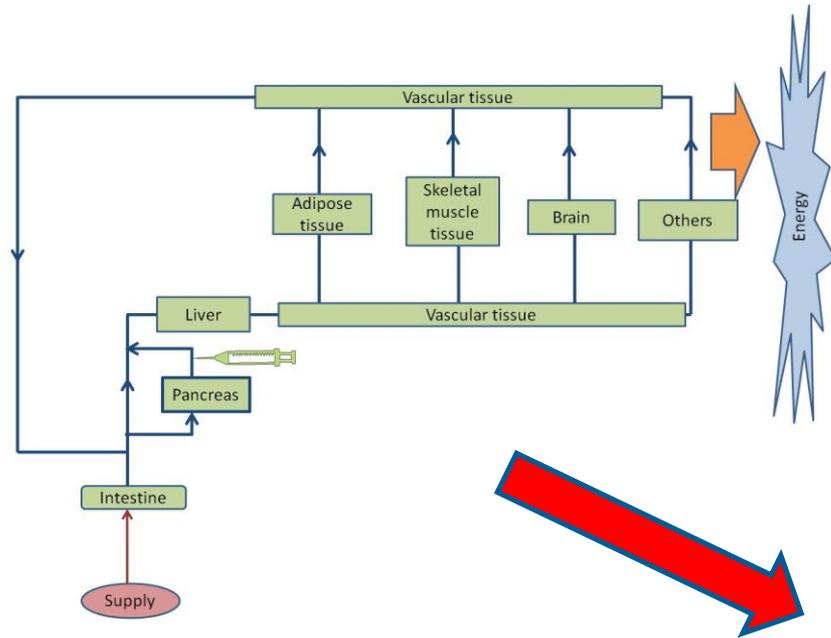
2 flow chambers

## Assays:

TEER, Alamar, LDH, vWF, cytokines, albumin, ZO-1, etc



# Connected Cell Culture



**Down-scaled and reduced  
to essential features of  
central metabolism**

